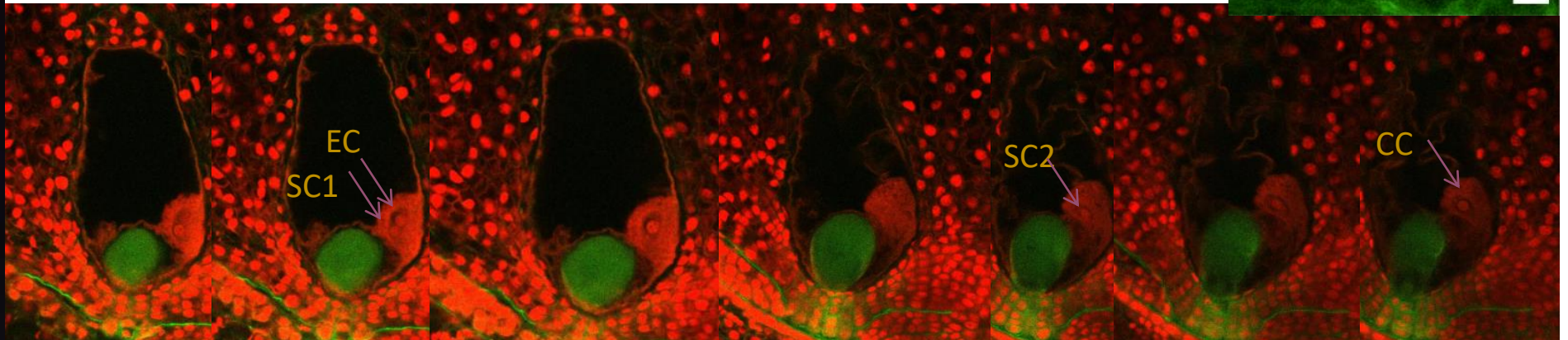
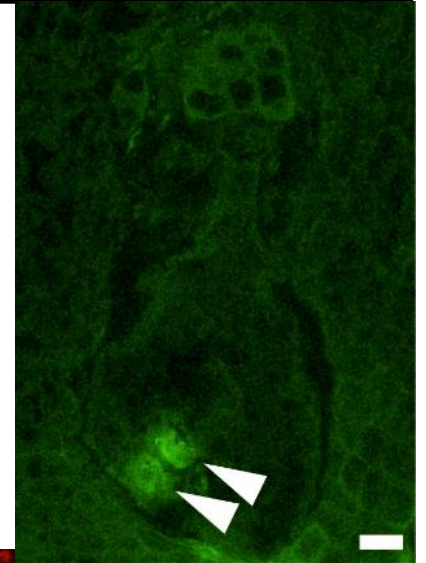
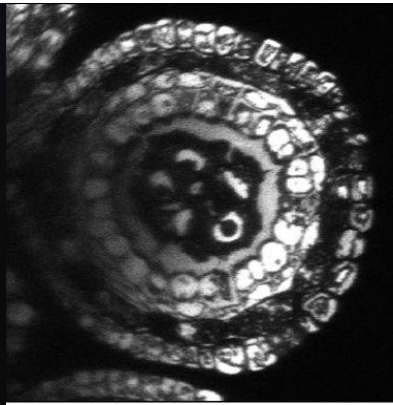


Reproductive Biology

Fertile Ground for New Breeding Technologies

Tim Kelliher, Principal Scientist in Reproductive Biology
Seeds Research, Syngenta Crop Protection

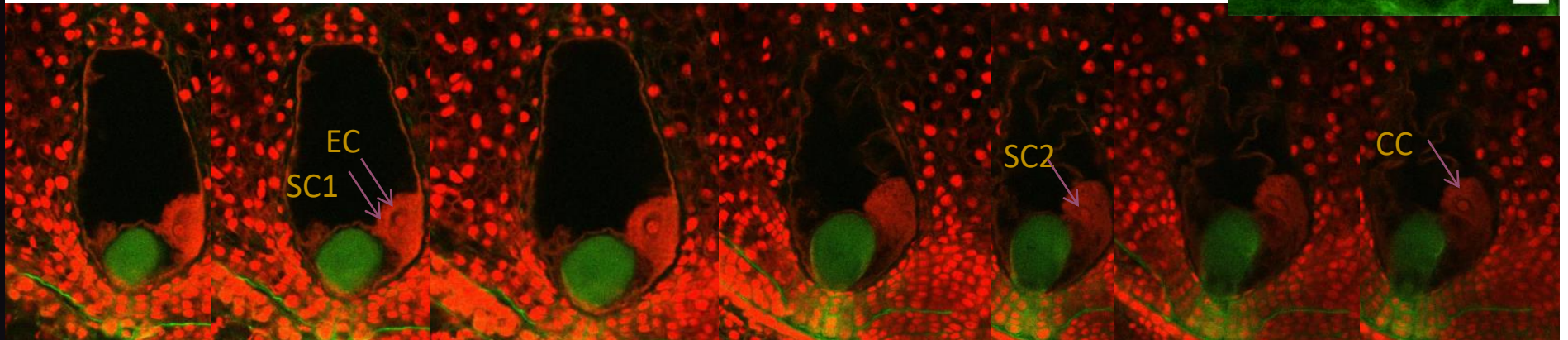
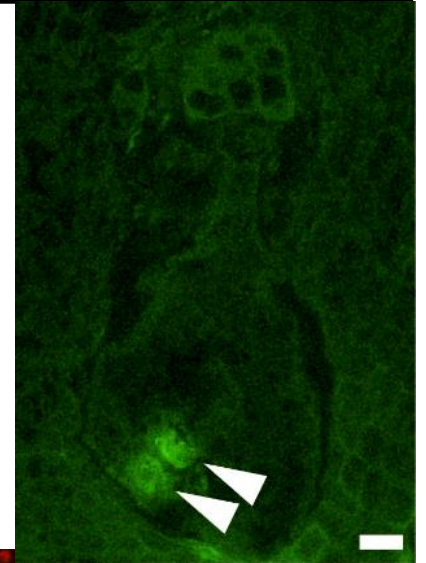
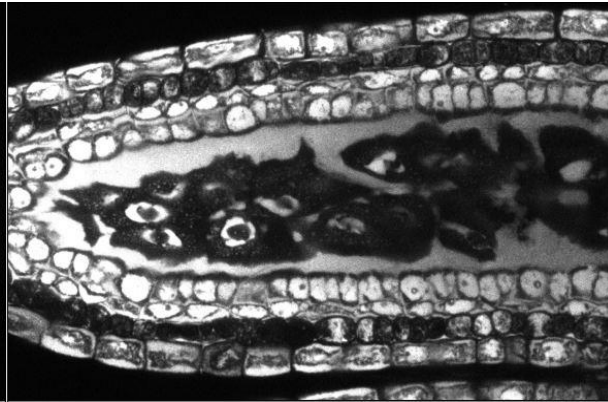
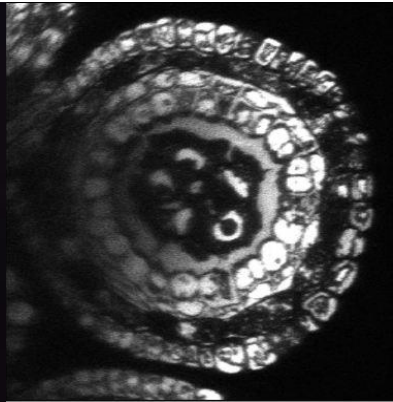
February 9th, 2018



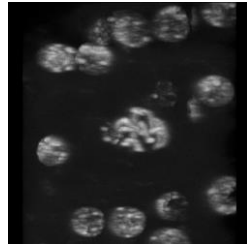
Reproductive Biology

Improving the sex life of corn

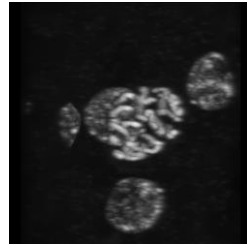
Tim Kelliher, Principal Scientist in Reproductive Biology
Seeds Research, Syngenta Crop Protection
February 9th, 2018



Haploids & doubled haploids

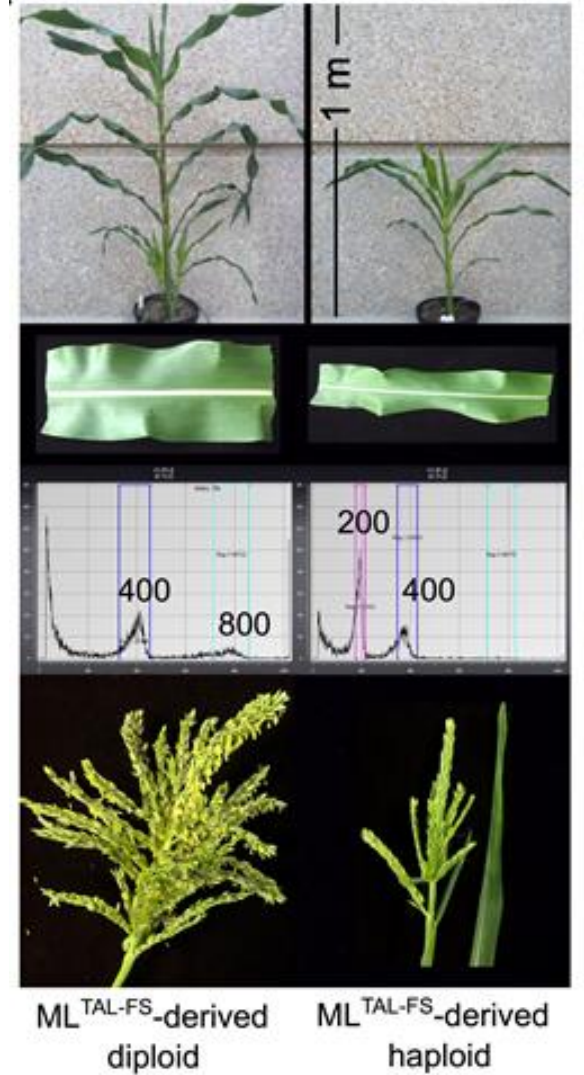


Haploid
10C



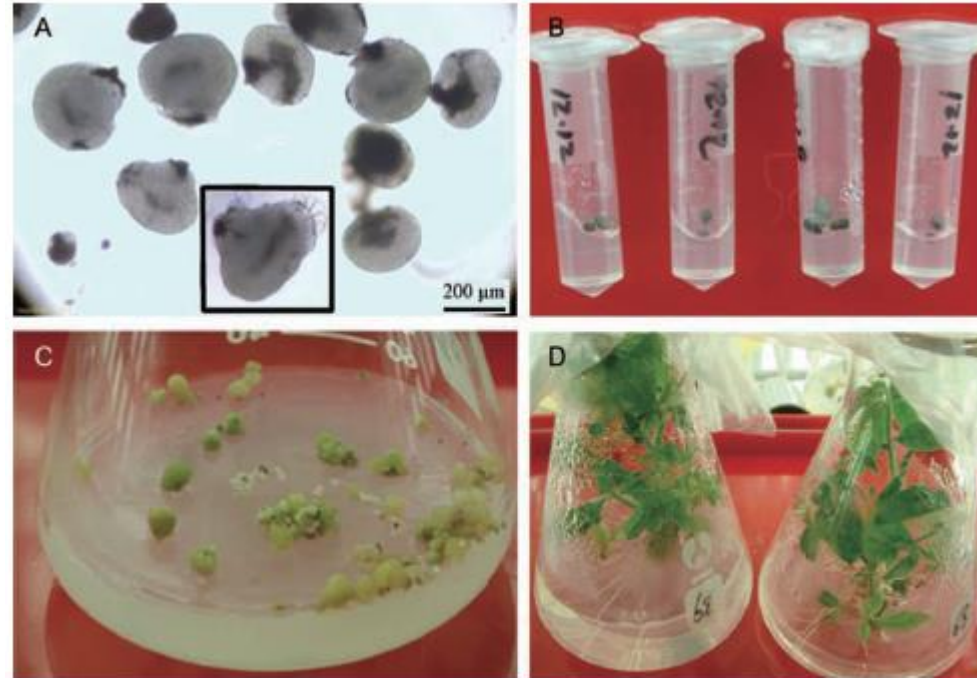
Diploid
20C

Haploid individuals have the gametic chromosome number (n) in their somatic cells



Haploid Induction Vary in Different in Different Crops

- Brassicas; many other species: Anther, microspore, or



fertilized (or)
endosperm

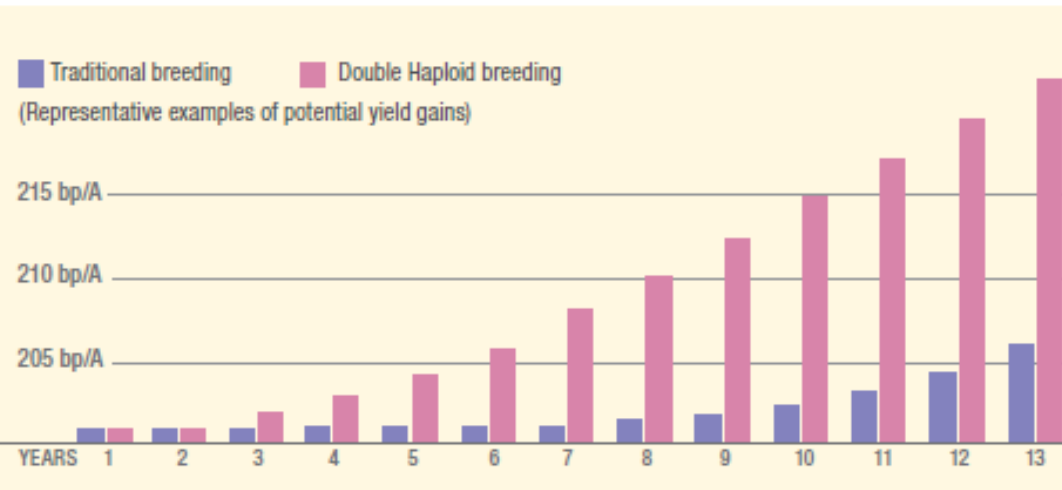
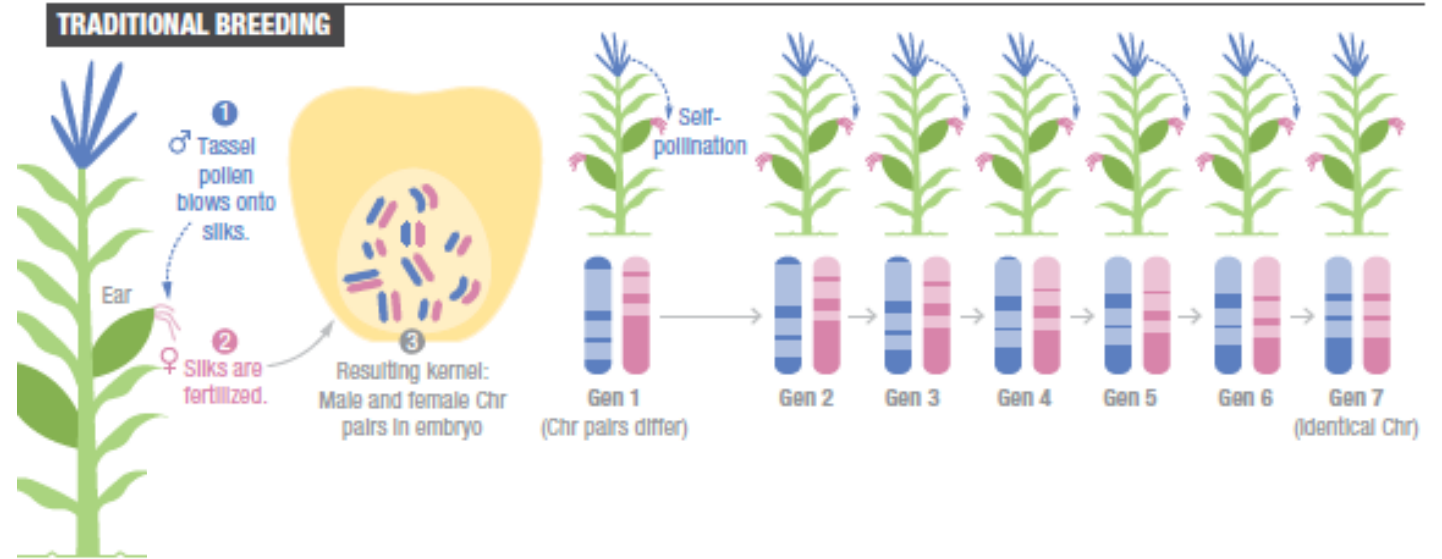
Ovule Pinar, Zhai, Euphytica 2015, 2014

Kasha, K.J. & Kao, K.N. High frequency haploid production in barley (*Hordeum vulgare* L.). *Nature* 225, 874-876 (1970)

Burke L. G., et al., Maternal haploids of *Nicotiana tabacum* L. from seed *Science* 206, 585 (1979)

- DH = Rapid production of inbred lines
 - Inbreds help us understand the connection between genes and traits
 - They are also needed to produce hybrids

The Benefits of Double Haploid Corn Breeding



1950s: Discovery of the first haploid inducing line (maize)

Vol. XCIII, No. 873

The American naturalist

November-December, 1959

LETTERS TO THE EDITORS

Correspondents alone are responsible for statements and opinions expressed. Letters are dated when received in the editorial office.

A LINE OF MAIZE WITH HIGH HAPLOID FREQUENCY

Chase (1949) has reported considerable variation in monoploid frequencies among different lines of maize, dependent upon both the maternal and paternal parents. The highest frequency found (0.688 per cent haploids from a particular single-cross hybrid crossed by a particular inbred pollen parent) is well above the average frequency of 0.111 per cent for all crosses used in the study. It is generally accepted that a haploid frequency of 0.1 per cent is usual.

A genetic inbred with an unusually high frequency of haploids has been found. Data accumulated over several years on self-pollinated progenies of an inbred designated as "stock 6" show 343 haploids in 10,616 observed plants, giving an over-all frequency of 3.23 per cent. Relatively few of these haploids have been verified by chromosome counts on root tips, as they are field-grown plants; however, the uniformity of stock 6 and the striking features of its haploids permit clear classification within the line with-



Advances in plant breeding



- Conventional Breeding
 - Introduce genes from the same or related species (wide cross introgression)
 - Induce variation (chemical mutagenesis, mutant screening)

RWK was obtained from Univ. Hohenheim; mapping initiated



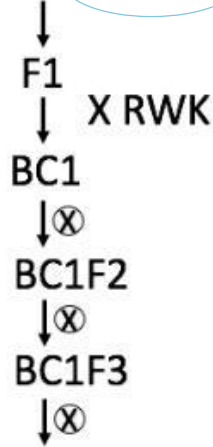
Satya Chintamanani



Brent Delzer

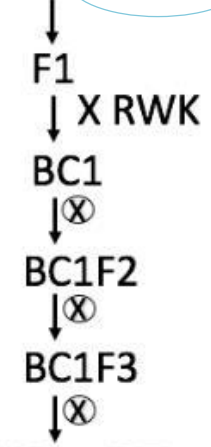
Initial Mapping

NP2391 X RWK



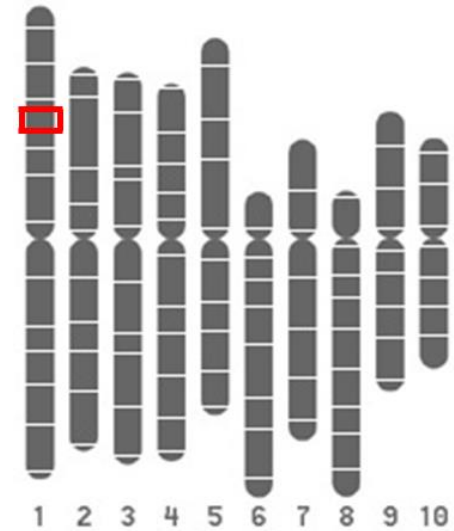
BC1F4 x 2 Testers

NP2460 X RWK



BC1F4 x 2 Testers

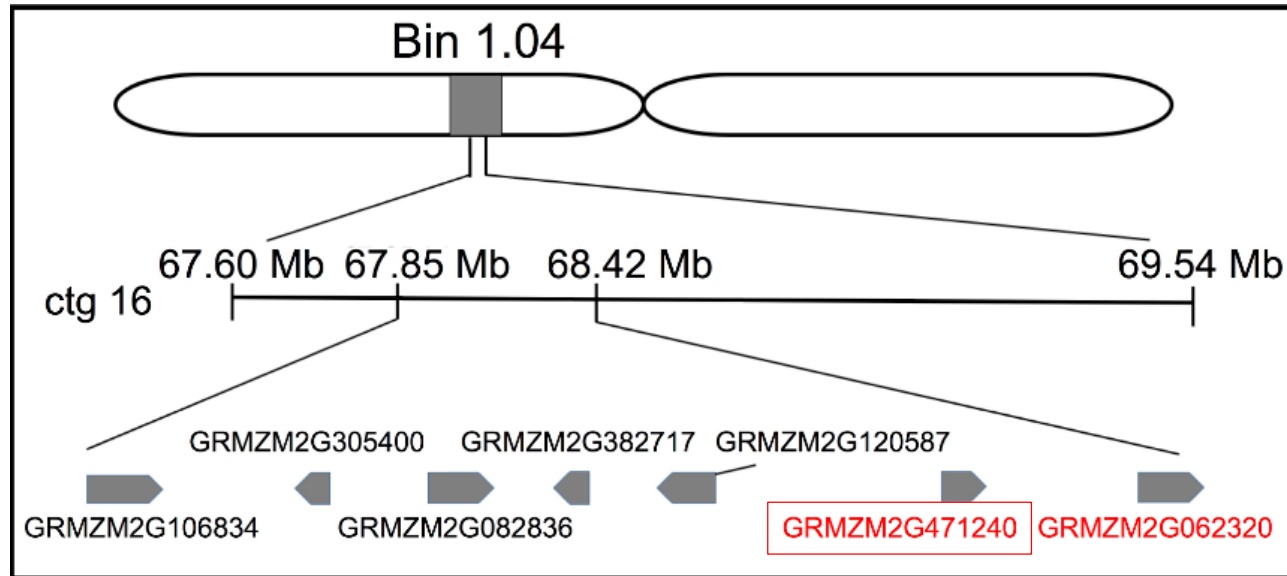
Mapped the QTL to Bin1.04



QTL =
Quantitative
Trait Locus

Qhir1 → 66%
of variation

2012: Seven genes in QTL sequenced in inducer (RWK, Stock6) and non-inducer lines (RWK-NIL, B73)

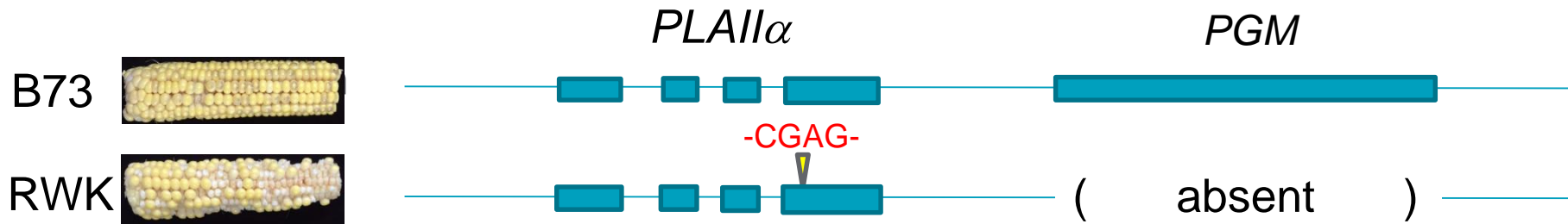


Research Triangle Park

- Mike Nuccio
- Bob Dietrich



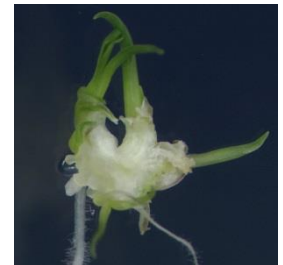
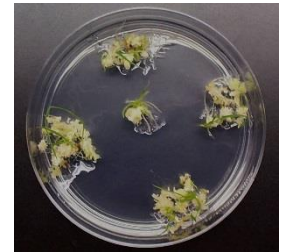
2013-2014: Complementation test, transgenic experiment



Pollen source	# ears	seed set	embryos	haploids	HIR
Control inducer	4	133/ear	255	26	10.20%
Control Inducer + $\frac{WT-PLA2}{WT-PLA2}$	16	357/ear	3047	7	0.23%



 *PLAI1 α*



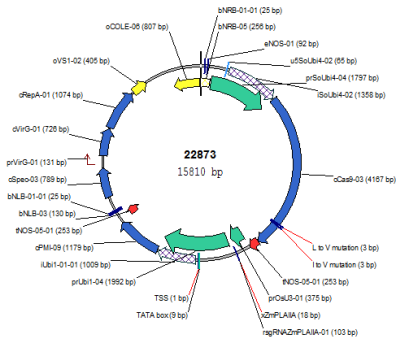
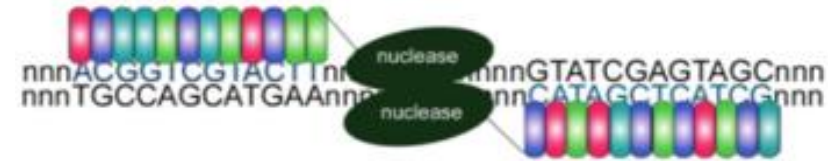
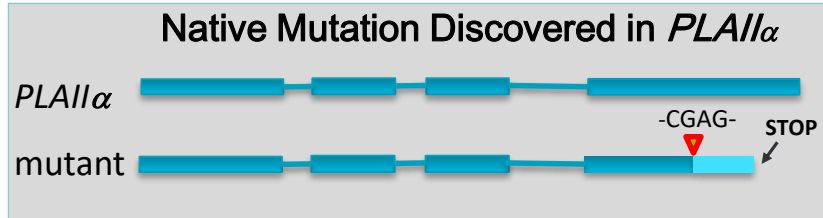
Michael Nuccio
Dakota Starr
Anna Prairie

Lee Richbourg
Heng Zhong
David Bradley



2015: TALEN editing provides the final proof

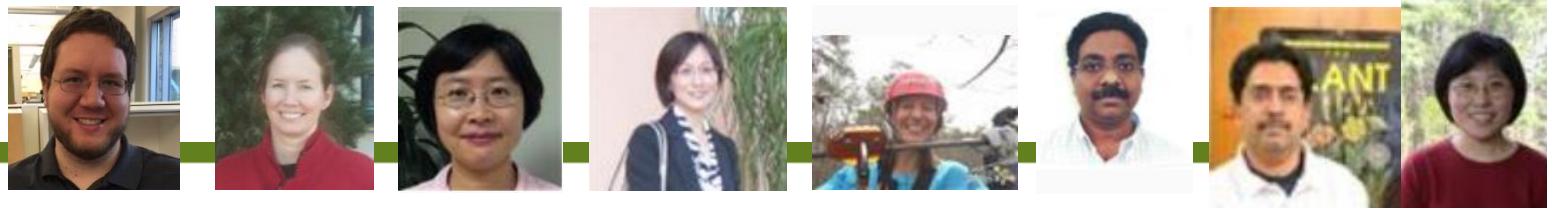
TALEN construct targeting native 4bp insertion site, induced several mutations

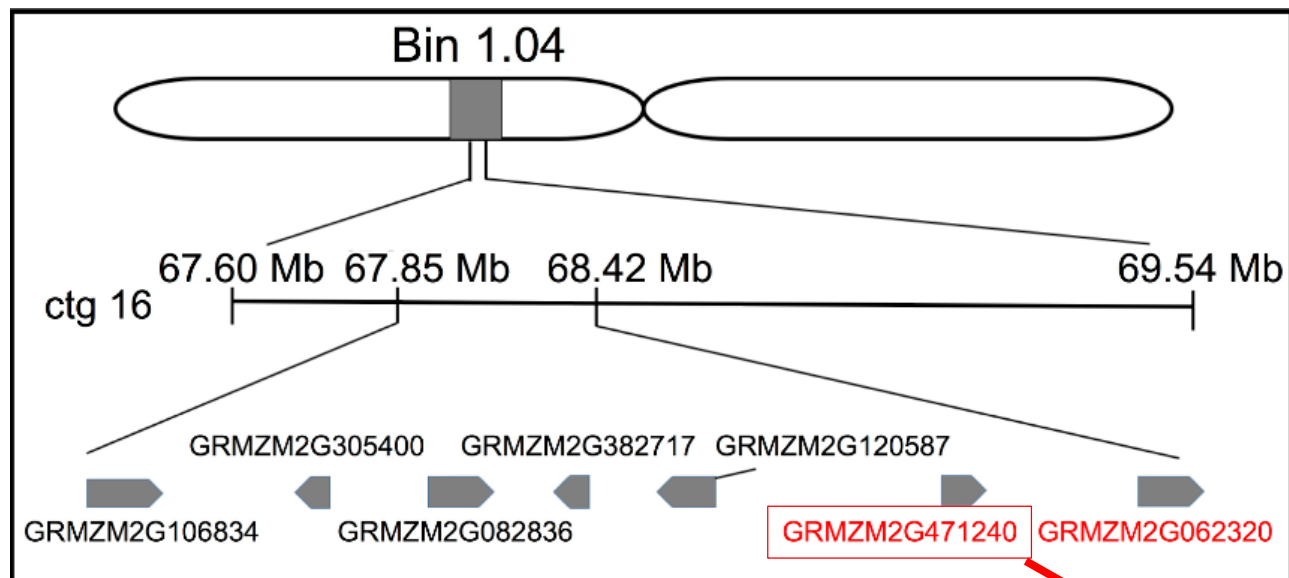


Pollen Source	Mutations	Ears	% Abort	Embryos	Putative Haploids	Confirmed Haploids	Induction Rate
AX	None	8	0.7%	1432	1	1	< 0.1%
Control inducer	4bp insertion	4	47.6%	531	56	54	10.2%
TAL-3954	-13bp; -28bp	4	44.10%	579	37	35	6.04%
TAL-3924	-8bp; -5bp	2	50.40%	128	18	16	12.50%
TAL-3932	-13bp	2	43.90%	169	18	15	8.88%
TAL-3317	-13bp	2	37.10%	343	19	19	5.54%
TAL-3303	-13bp	1	34.60%	176	7	7	3.98%
TAL-4108	-11bp; -5bp	4	40.10%	379	28	26	6.86%

Dakota Starr
 Jamie McCuiston
 Zhongying Chen
 Shujie Dong

Tara Liebler
 Sam Nalapalli
 Siva Elumalai
 Wenling Wang



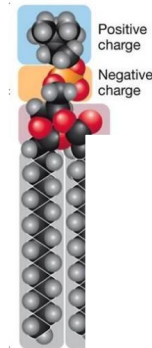
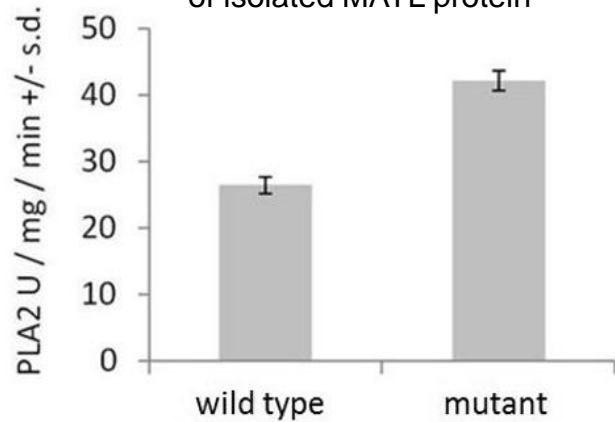


PLAIIα renamed *MATRILINEAL*

How does MATRILINEAL induce haploids?

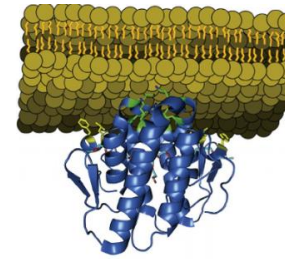
Phospholipases cleave fatty acid chains off of membrane phospholipids

In vitro phospholipid cleavage assay of isolated MATL protein

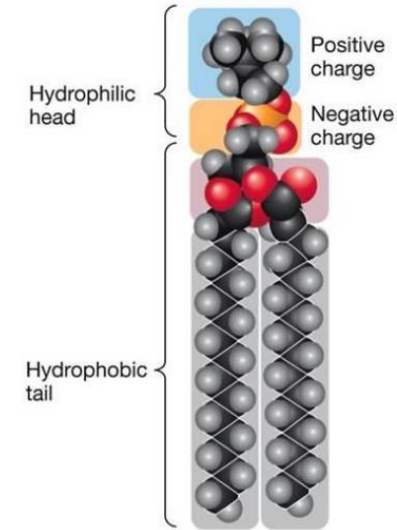


Lyso-phospholipid

→ signaling, membrane remodeling



MATL



Free fatty acid → prostaglandins (animals)

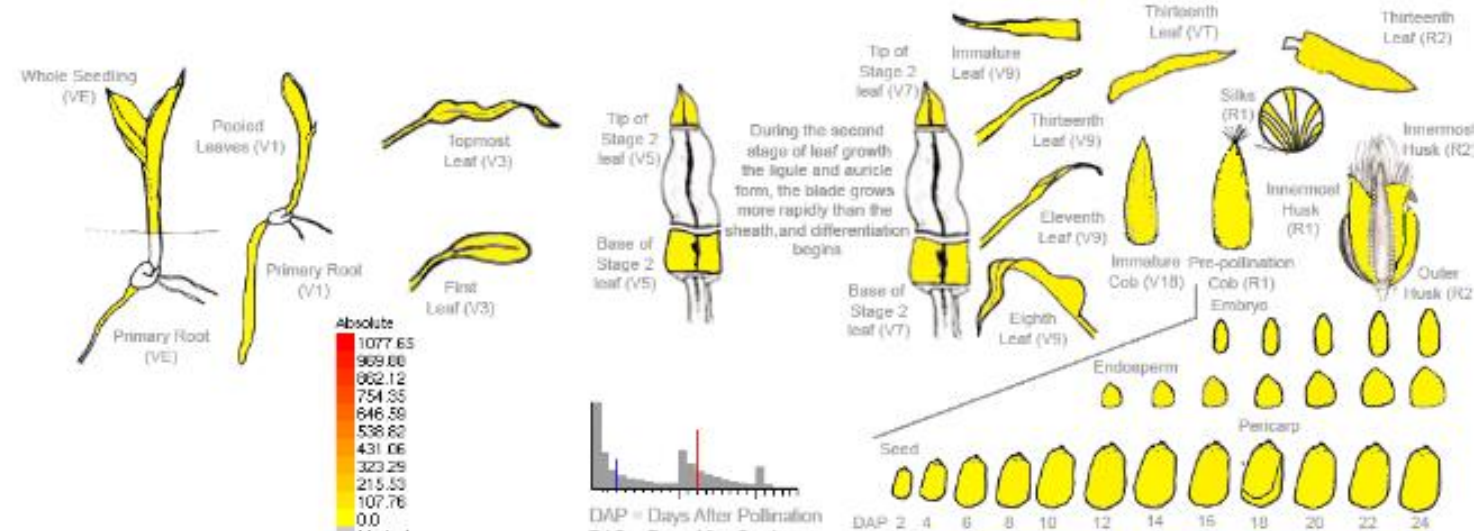
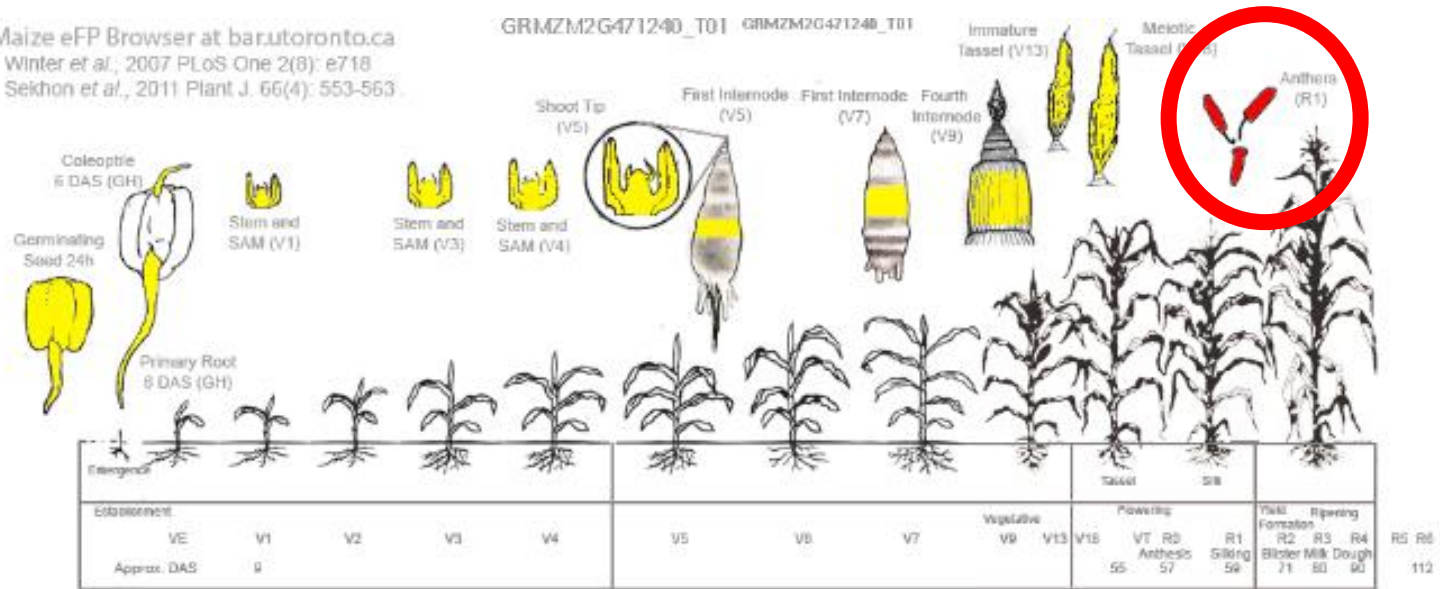
oxylipins (plants)



How does MATRILINEAL induce haploids?

Expression pattern of *Matrilineal* RNA – anthers only!

Maize eFP Browser at bar.utoronto.ca
 Winter et al., 2007 PLoS One 2(8): e718
 Sekhon et al., 2011 Plant J. 66(4): 553-563.



Formation of the male gametophyte (pollen)

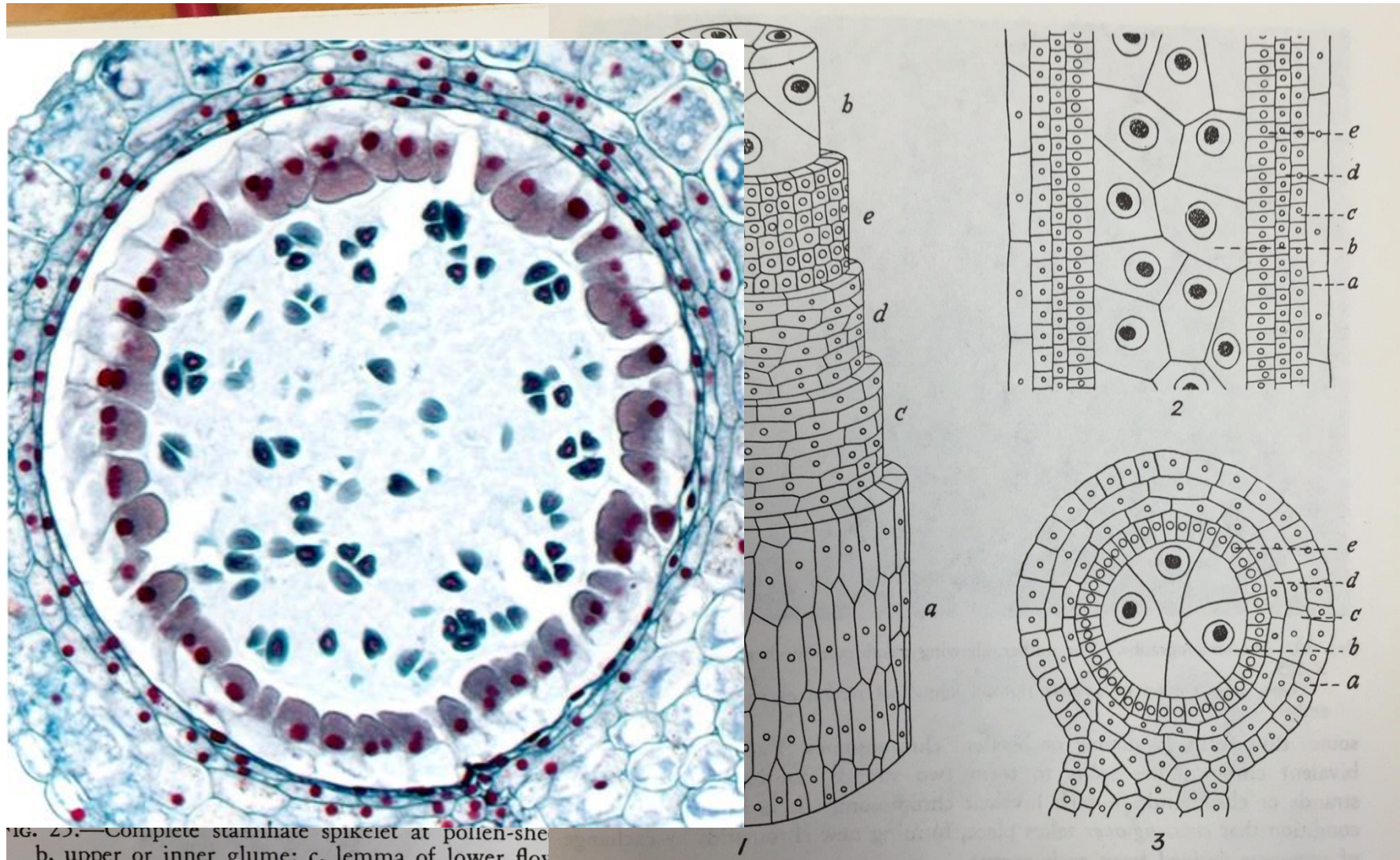


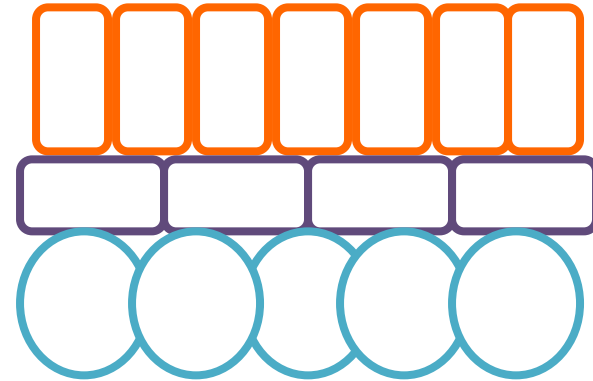
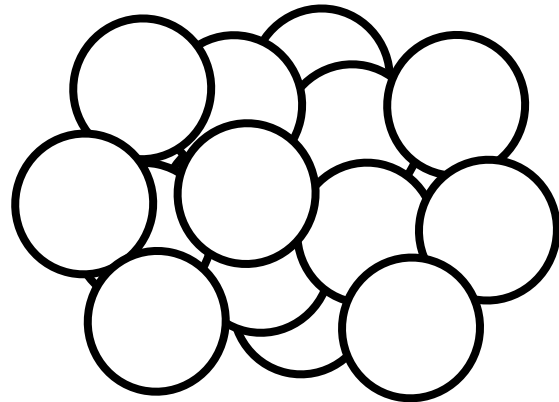
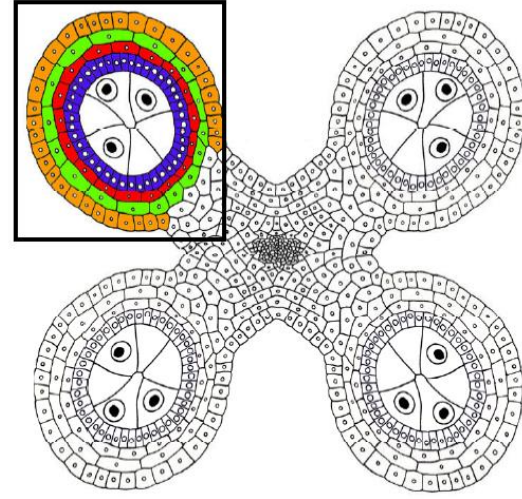
FIG. 25.—Complete staminate spikelet at pollen-stage.
b, upper or inner glume; c, lemma of lower floret.

Theodore Kiesselbach, 1949 The structure and reproduction of corn
University of Nebraska, Agricultural Experimental Station

fate acquisition

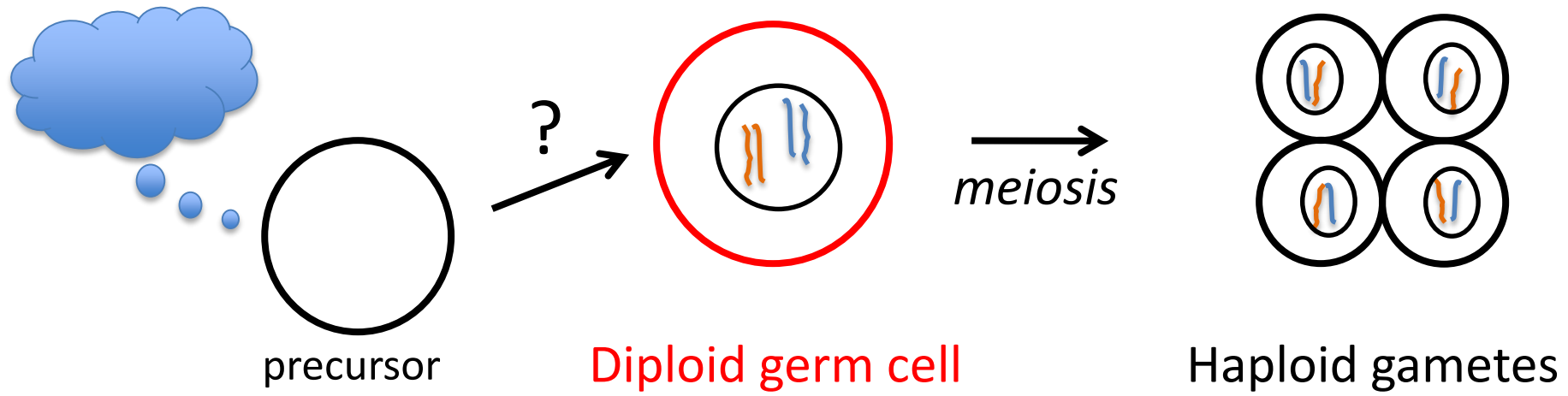


differentiation



*tissue composition
and arrangement*

Cell walls → Fixed positions → get to know your neighbors!



Germ cells: the ultimate totipotent cell
All others (somatic): “dead end” lineages

Germ cell specification in animals and plants



early sequestration, quiescence

induction by somatic cells

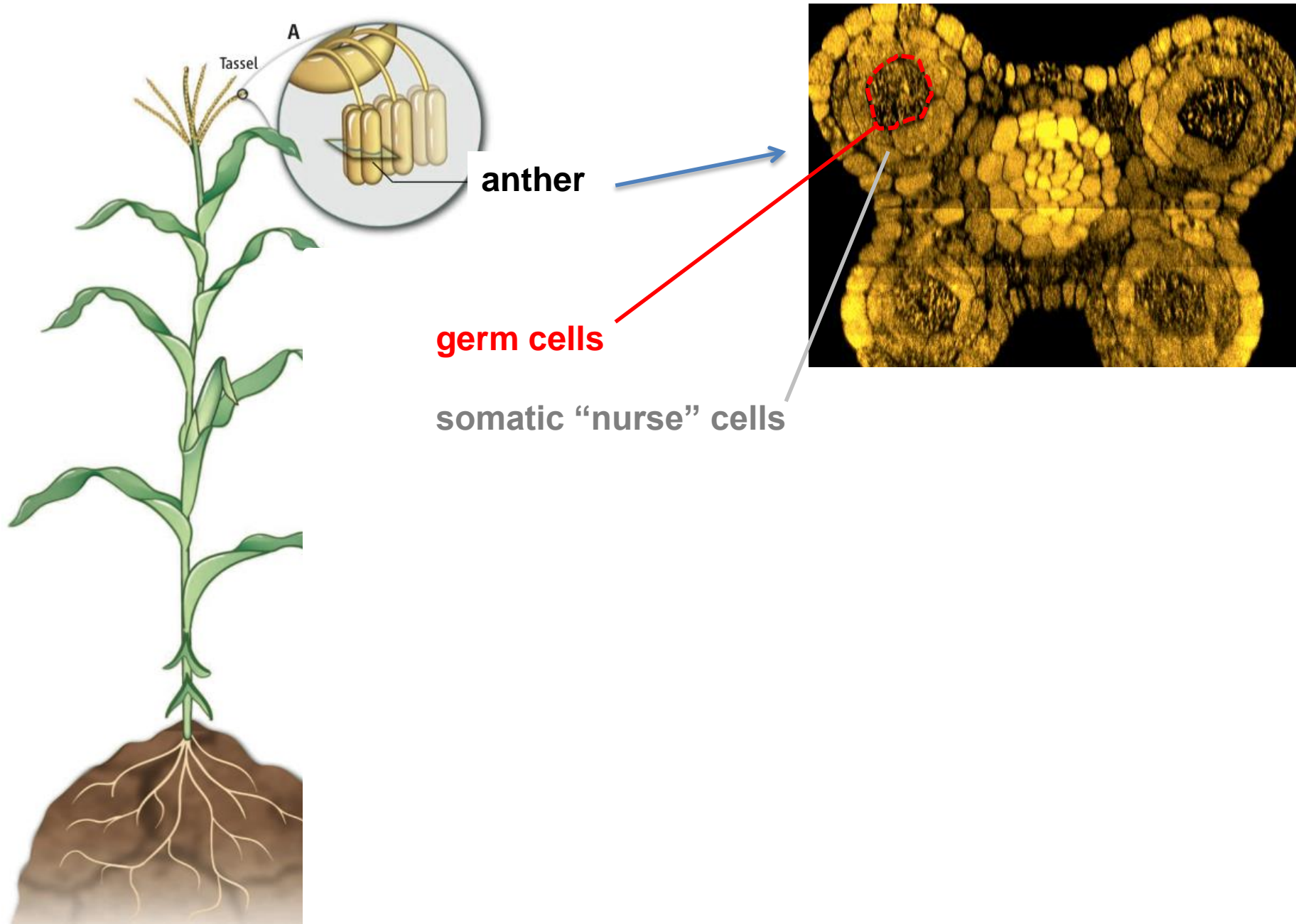
continuous production



late specification

need for protection from DNA damage

When and where do germ cells arise?



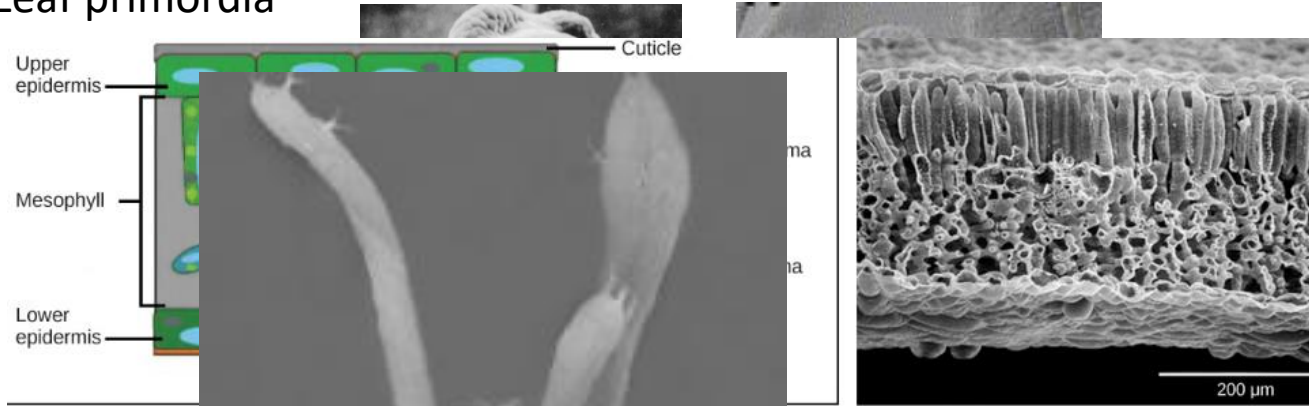
Leaves

vs.

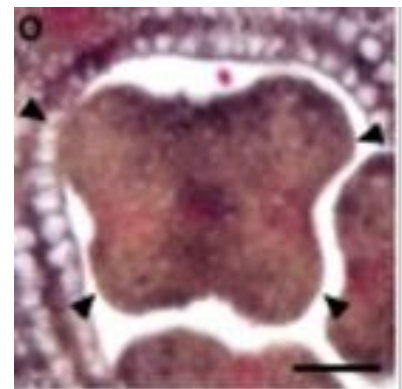
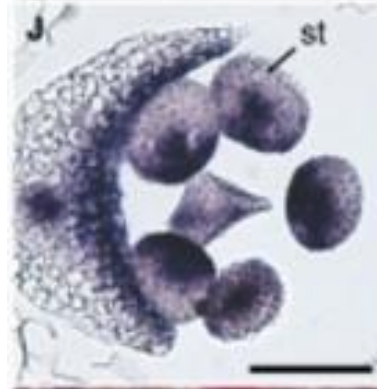
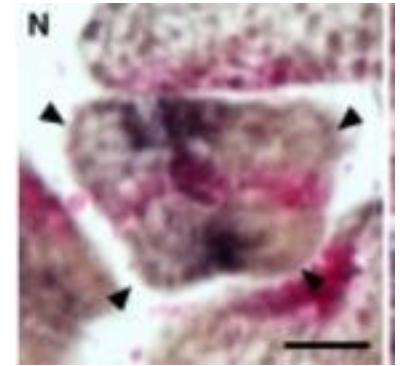
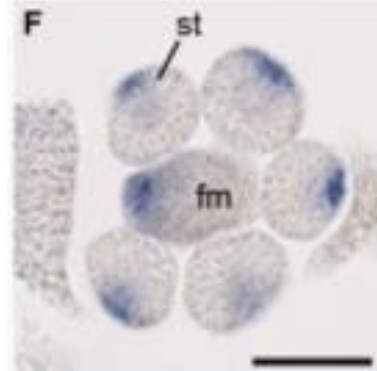
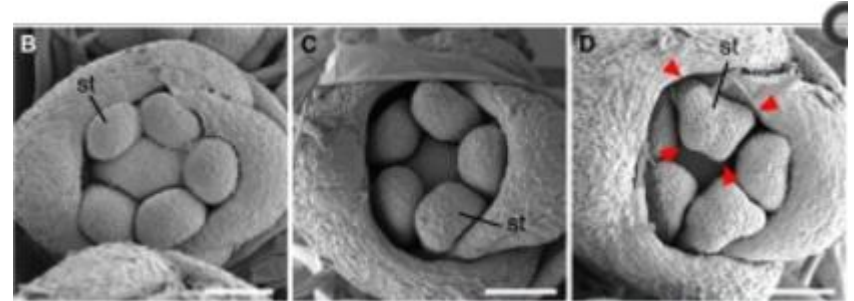
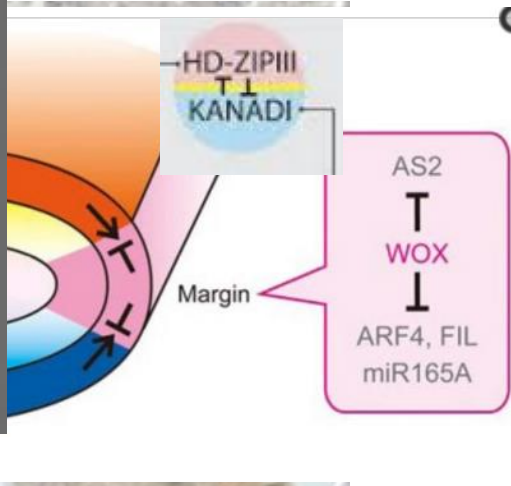
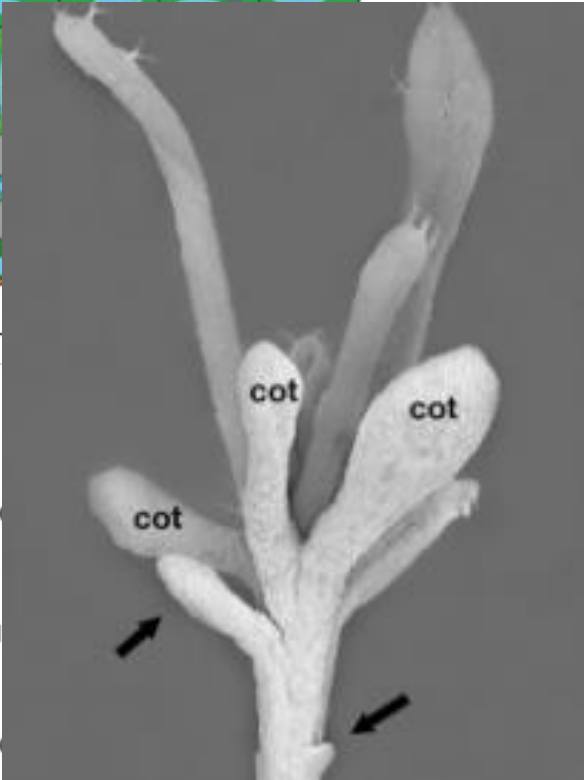
Anthers

Shoot apical meristem

Leaf primordia

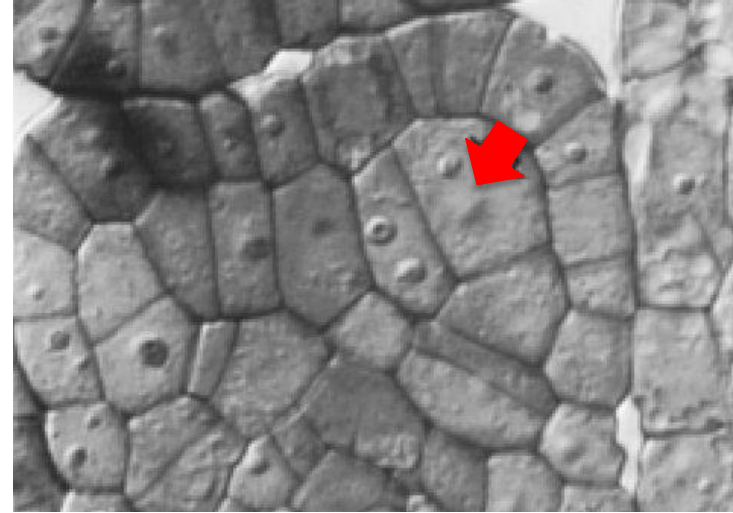


Adaxial ↑
Abaxial ↓

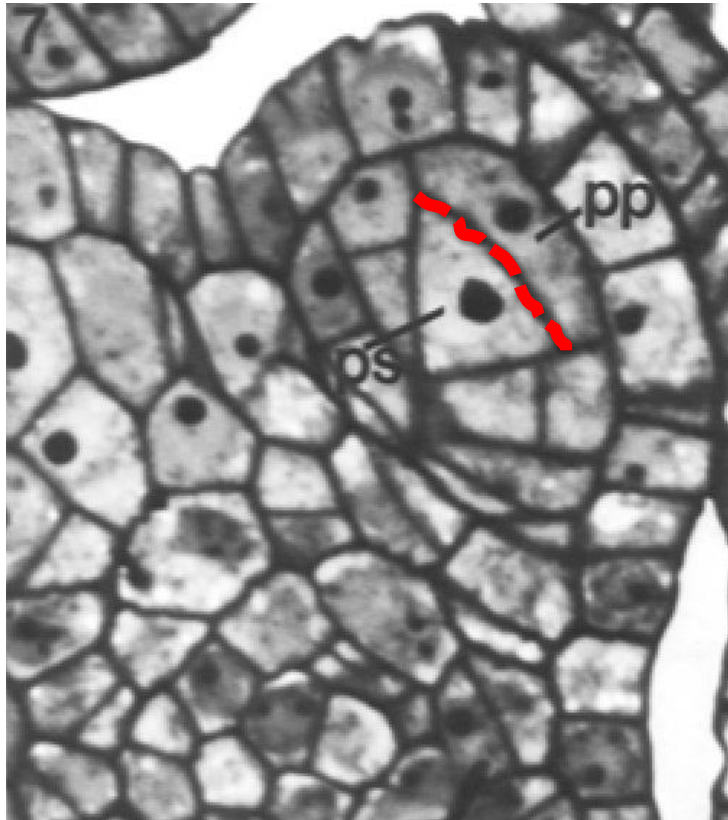


TEXTBOOK VIEW: lineage defines germinal fate
asymmetric cell division (ACD) is the mechanism

hypodermal cell



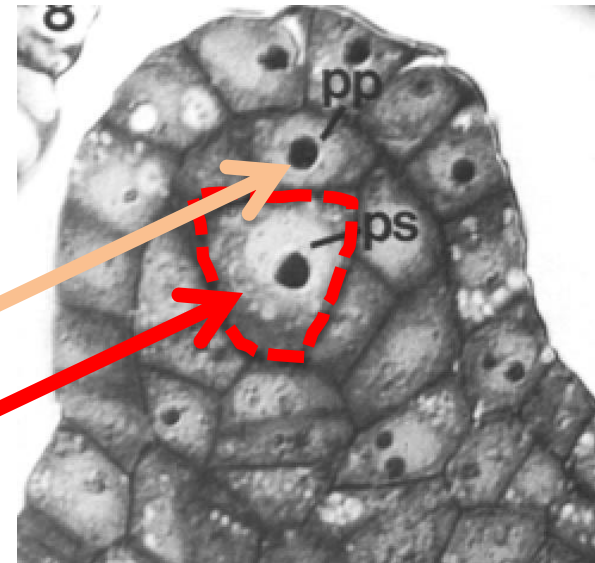
Raghavan, V., 1988. *Am J. Bot.* 75(2) (rice)



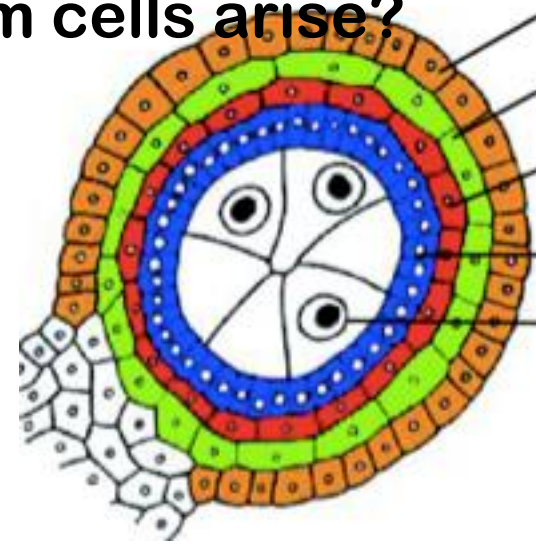
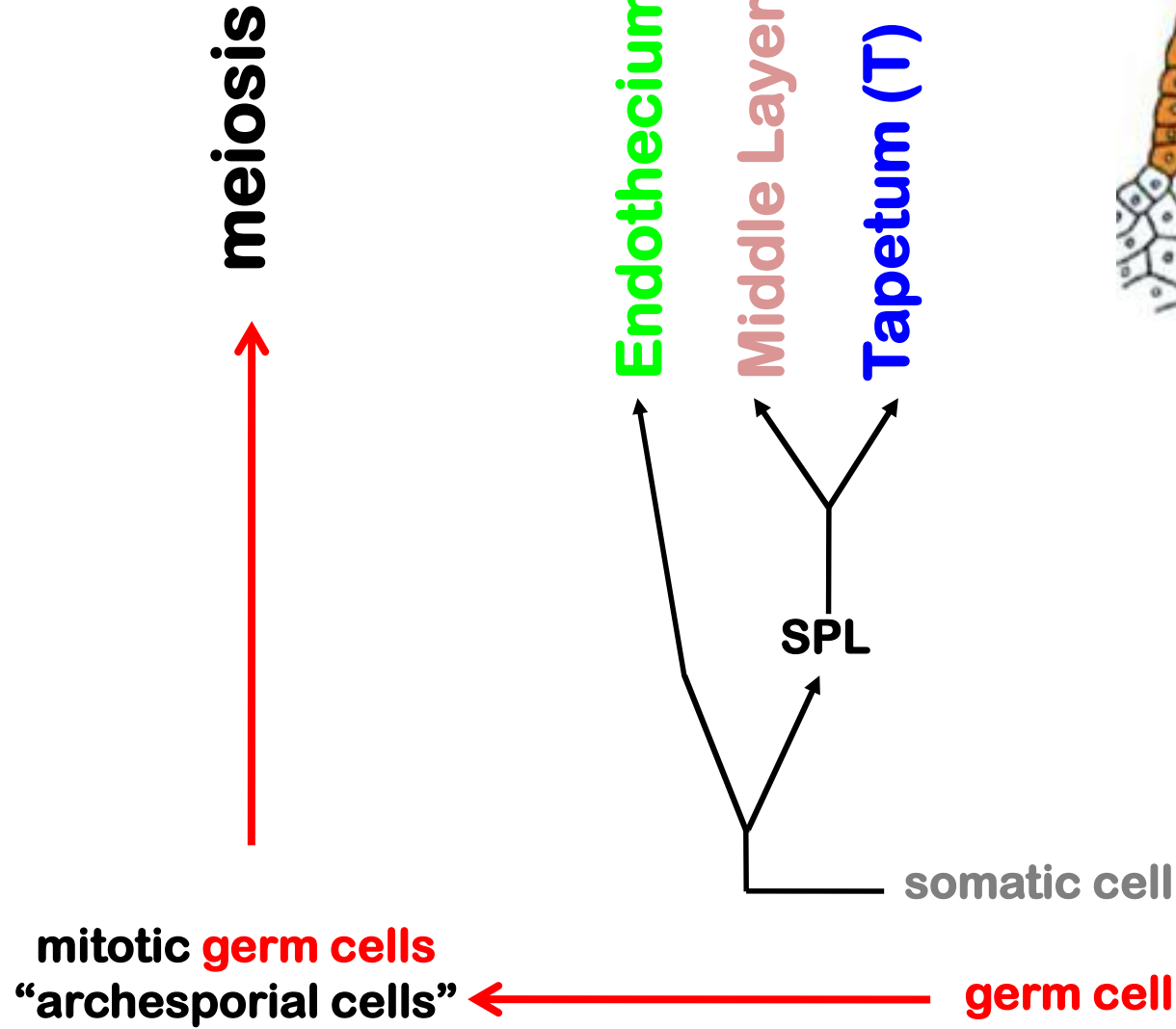
undergoes ACD

somatic cell

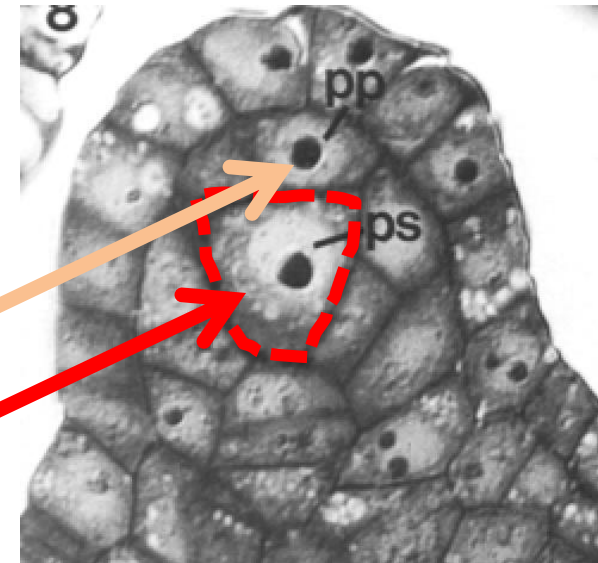
germ cell



When and where do germ cells arise?

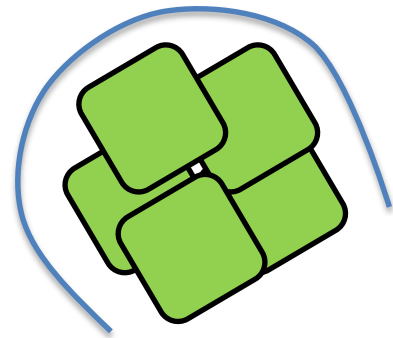
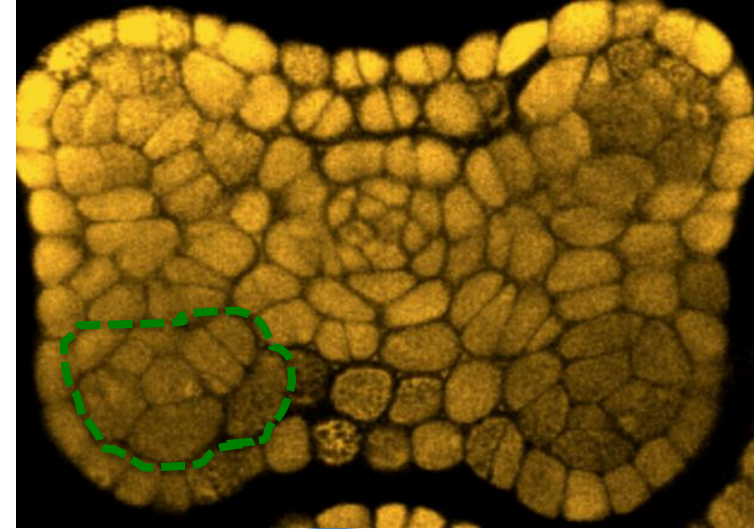


Ma, et al., 2008. *Genome Biology*

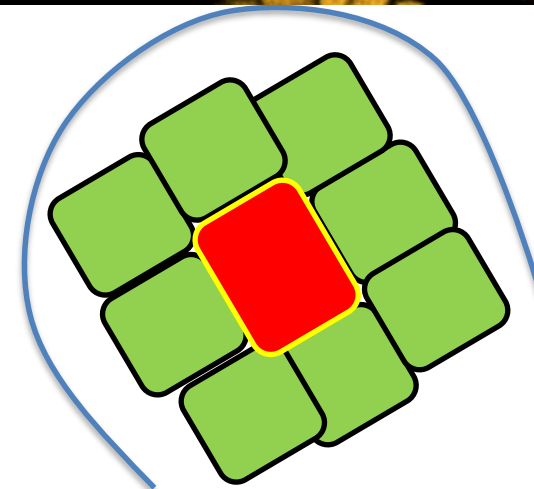


When and where do germ cells arise?

~ 30-40 morphologically equivalent **L2-d cells** per lobe

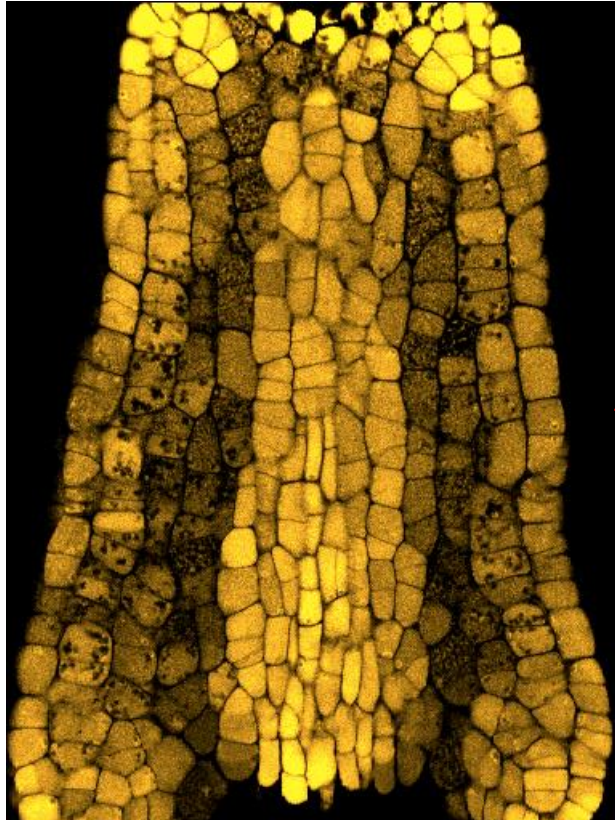
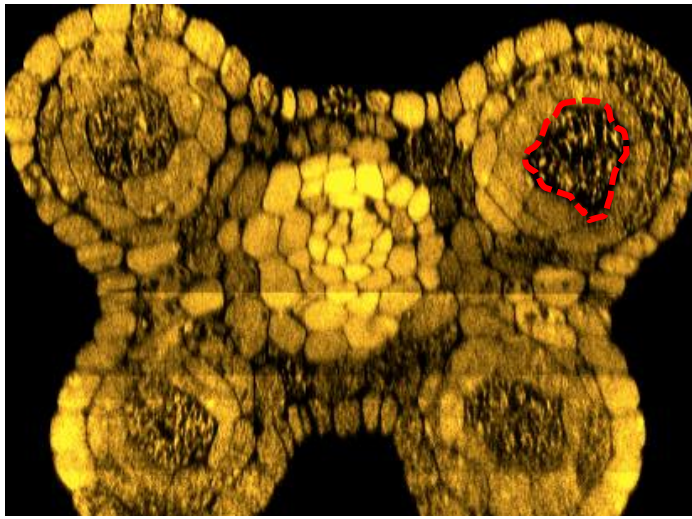


L2-d cells



AR (germ line)
+ L2-d cells (somatic niche)

NEW APPROACH: confocal microscopy

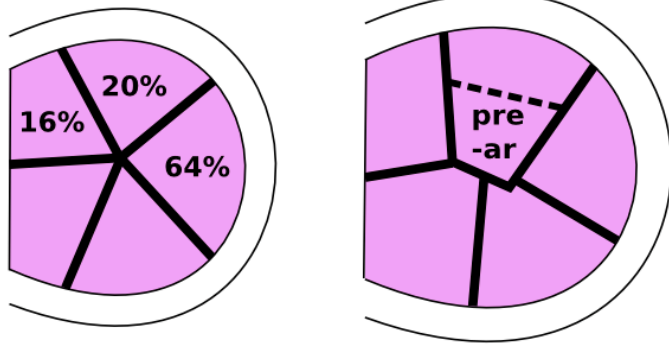


**Characteristics
of germ cells**

- Darker stain
- Rounded shape
- Central position

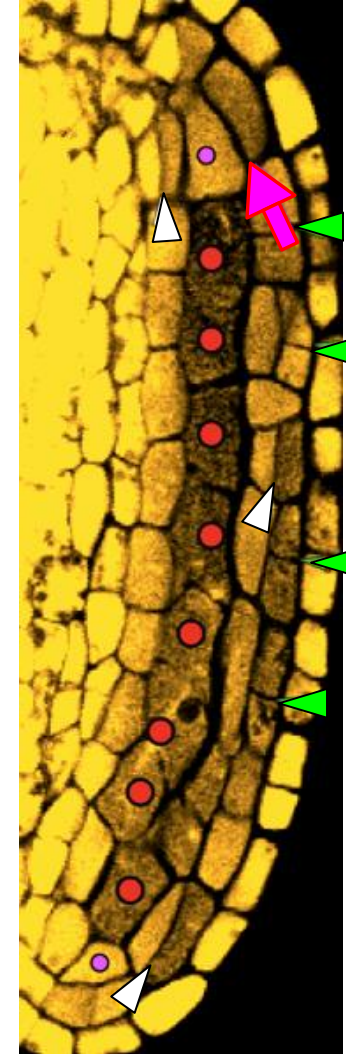
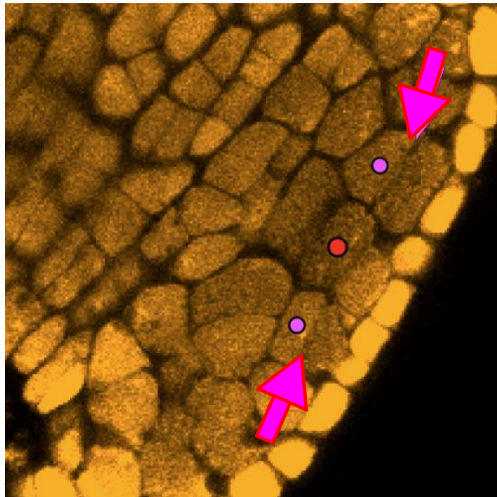


When and where do germ cells arise?



- multiple progenitors
- center more advanced than base and tip

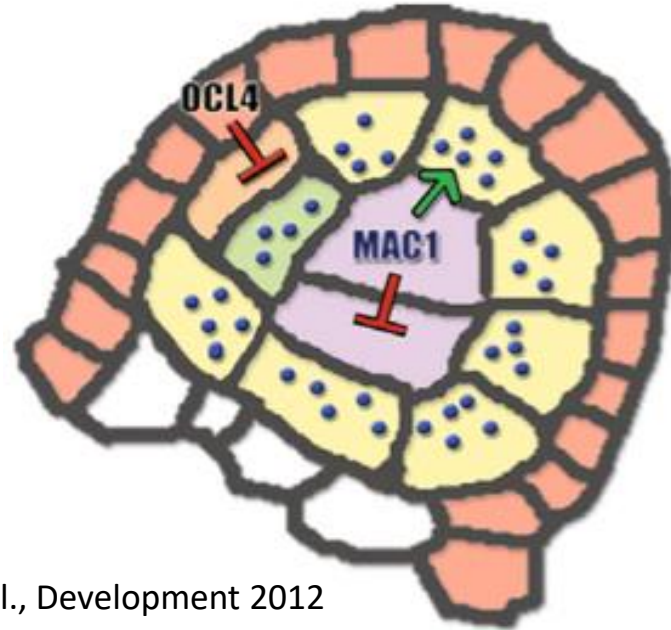
● presumptive germ cell
● germ cell (AR cell)



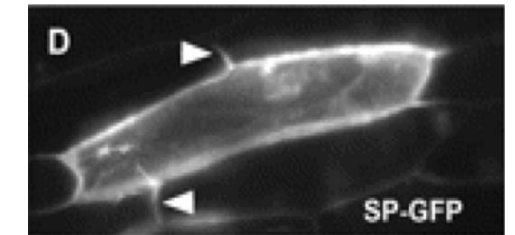
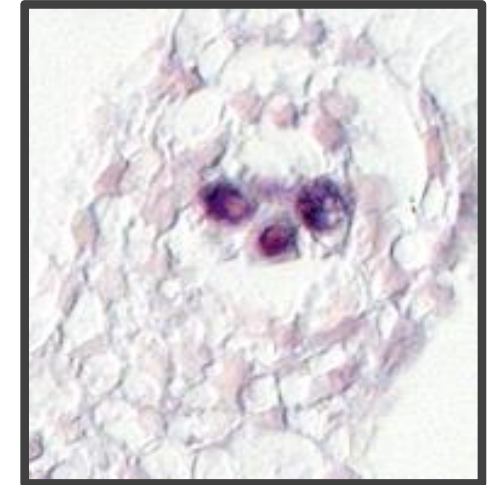
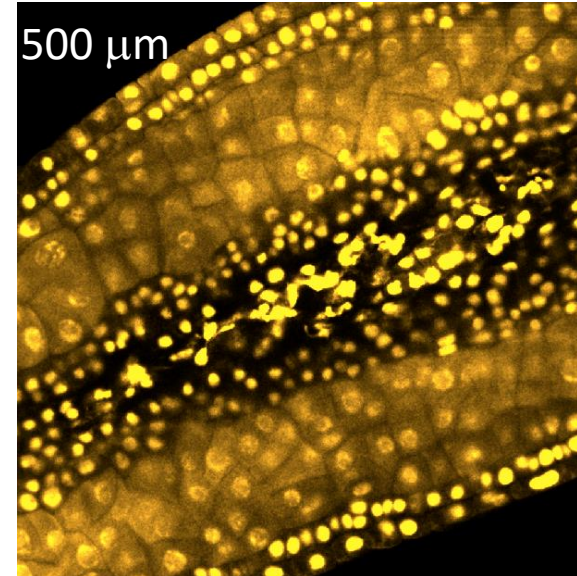
Hypoxia as
Positional cue

AR express MAC1, a secreted ligand

fertile



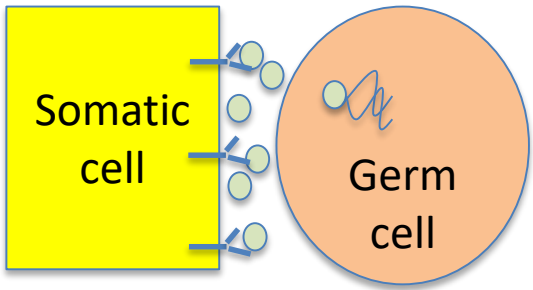
mac1



Wang, et al., Development 2012

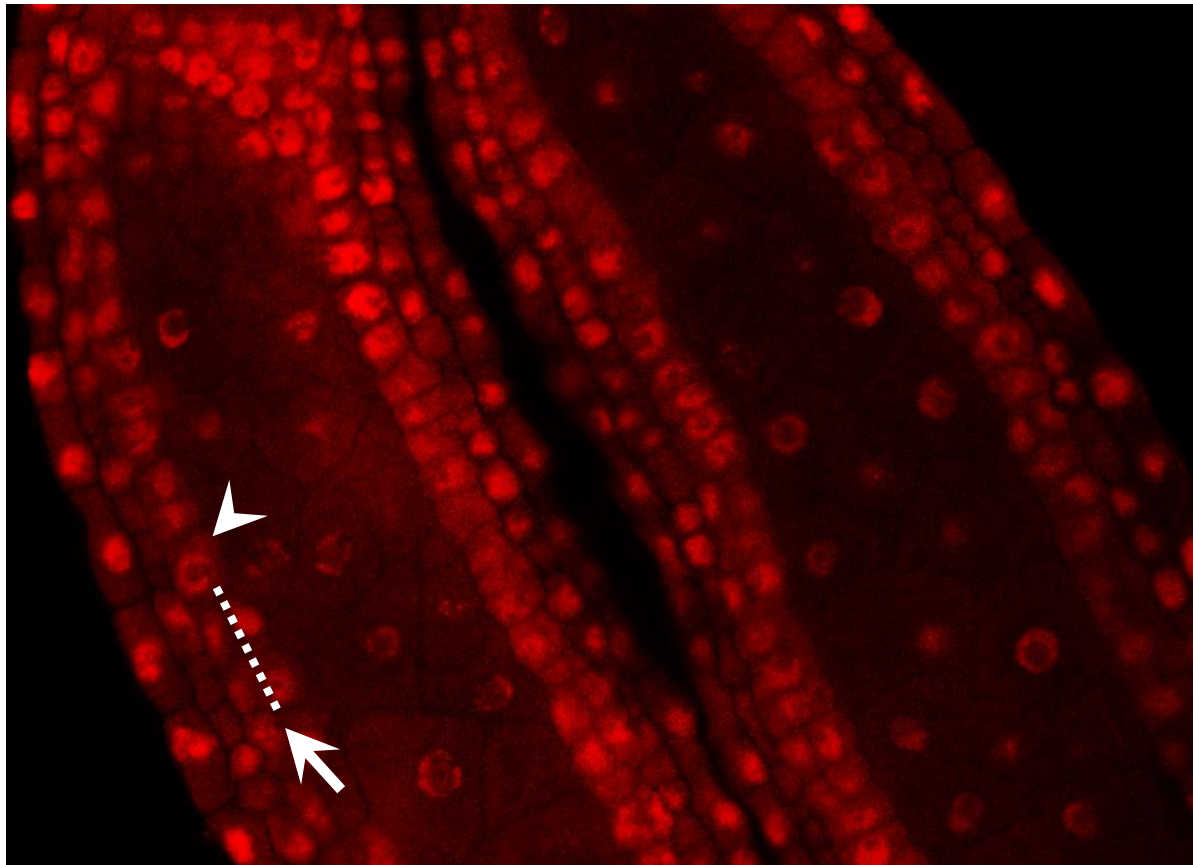
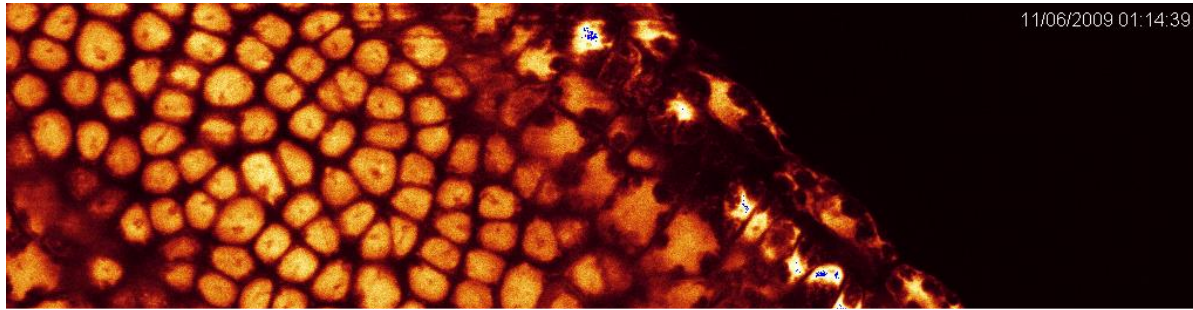
**Excess germ cells
Faulty somatic development**

● MAC1 small Peptide (CLV3-like)

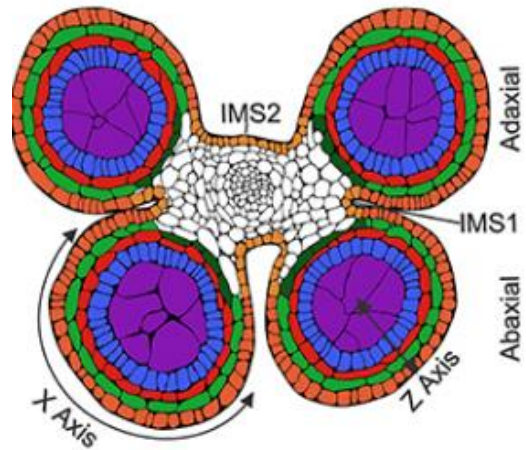


— EMS1 LRR-RLK receptor

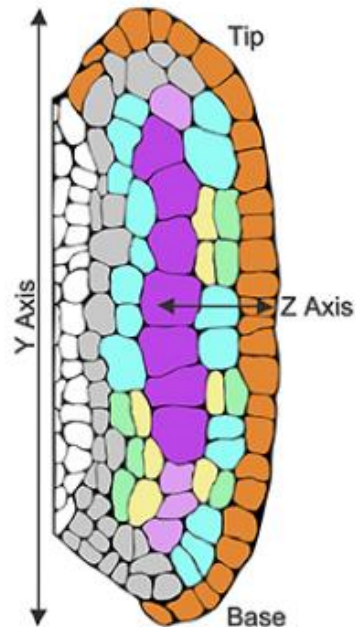
Receptor mutant has never been found → gene editing!











A Transverse: Stage 8



C Longitudinal: Stage 3



B

		Anther length (μm) or [Stamen length (μm)]			
		Arabidopsis	Maize	Rice	
	L1-derived (L1-d) L2-derived (L2-d)	1. Pluripotent primordium	<10	30-120	50-100
	Primary Parietal Cell (PPC) Archesporial-Specified	2. AR arise, triggering PPC differentiation	<20	120-180	100-150
	Epidermis (EPI) Endothecium-Specified Secondary Parietal-Specified Archesporial (AR) OCL4-Dependent Epidermis Endothecium (EN) Secondary Parietal Cell (SPC)	3. EN and SPC forming	<30	180-280	150-250
	Non-Subepidermal Endothecium	4. EN and SPC complete; anticlinal division adds to girth	35*	280-500	275*
	Middle Layer-Specified Tapetum-Specified	5. Bipotent SPC divisions start to make ML and TAP	40	500	300
		6. SPC periclinal divisions and anticlinal growth	70*	500-700	350*
	Middle Layer (ML) Tapetum (TAP)	7. Cell differentiation and final SPC periclinal divisions	100*	700	400
	Pollen Mother Cell (PMC)	8. Lobe growth and AR mature to PMC	120	1000-1200	400*
		Meiosis Starts	240	1500	400-450

Formation of the male gametophyte (pollen)

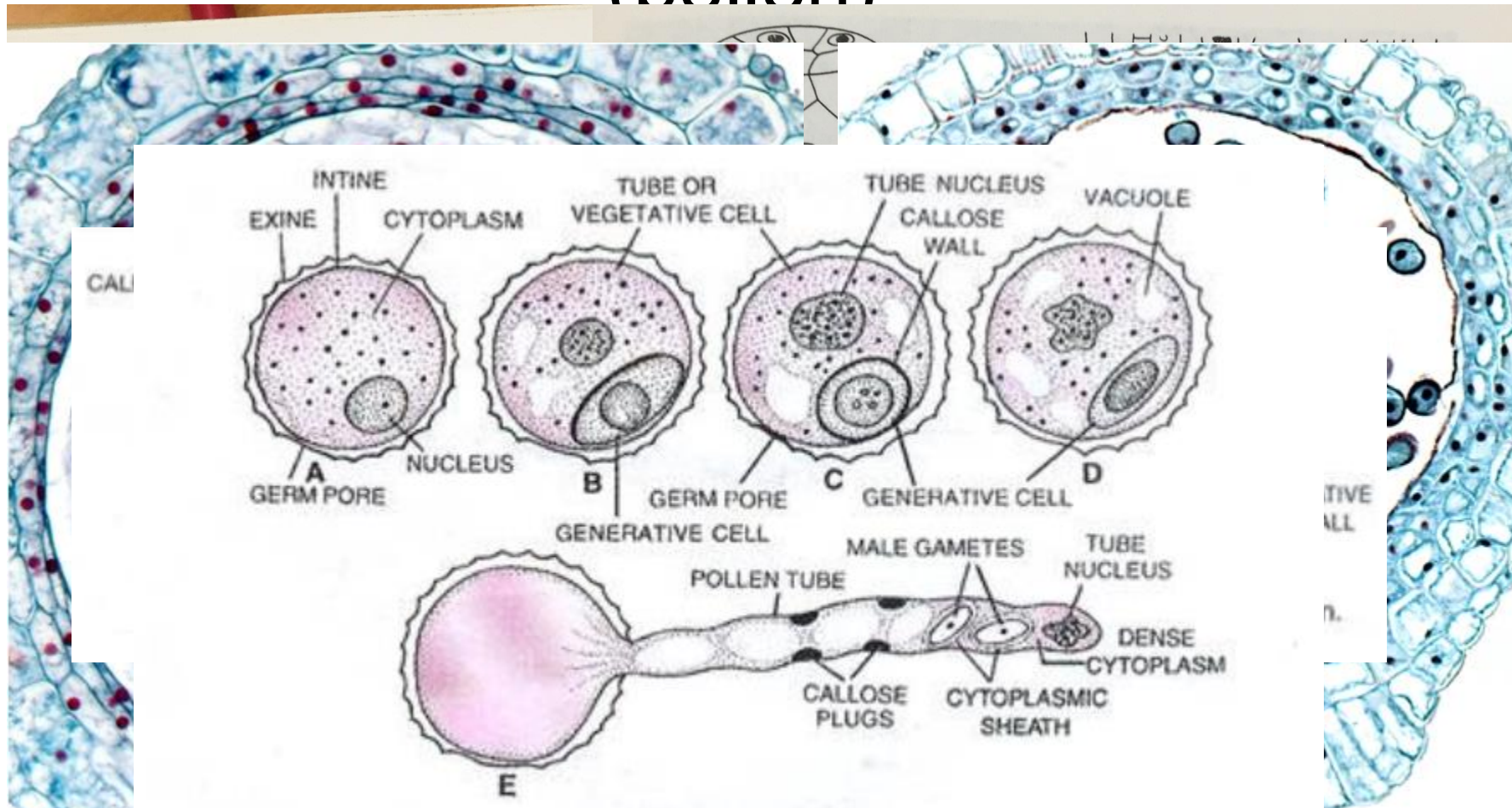


Fig. 2.9. Germination of pollen grain and formation of male gametophyte in an angiosperm.

FIG. 25.—Complete staminate spikelet at pollen-sheath
 b. upper or inner glume; c. lemma of lower floret

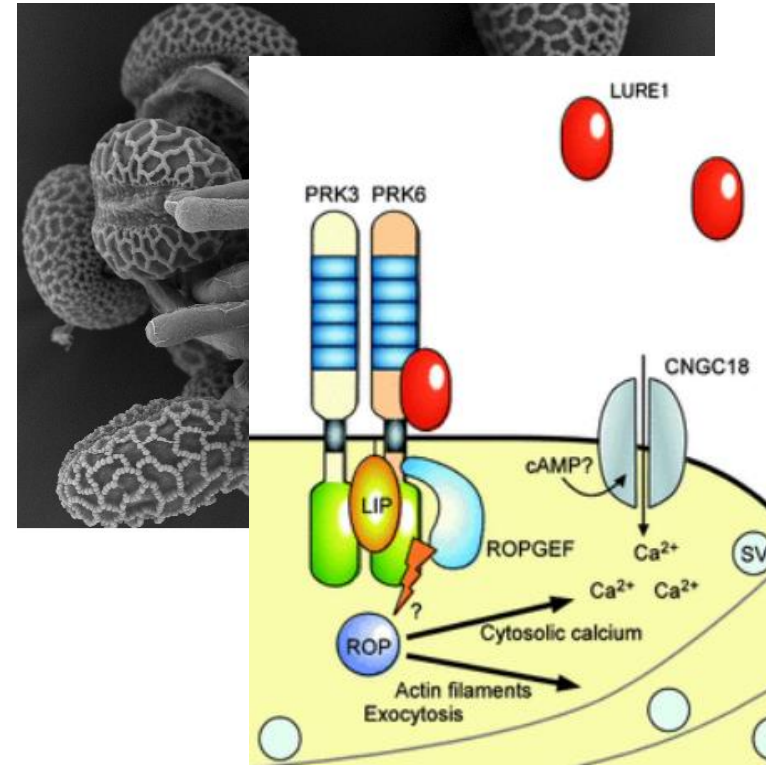
Theodore Kiesselbach, 1949 The structure and reproduction of corn

Formation of the female gametophyte (embryo sac)



Defensin-like polypeptide LUREs are pollen tube attractants secreted from synergid cells.

Tetsuya Higashiyama,
Nagoya Univ.



PRK3/6 are the
pollen tube tip-
localized receptors
of the LUREs

[In vitro pollination, Torenia](#)

[Torenia - demonstrating guidance cues](#)

[Guidance cues alternative link \(youtube\)](#)

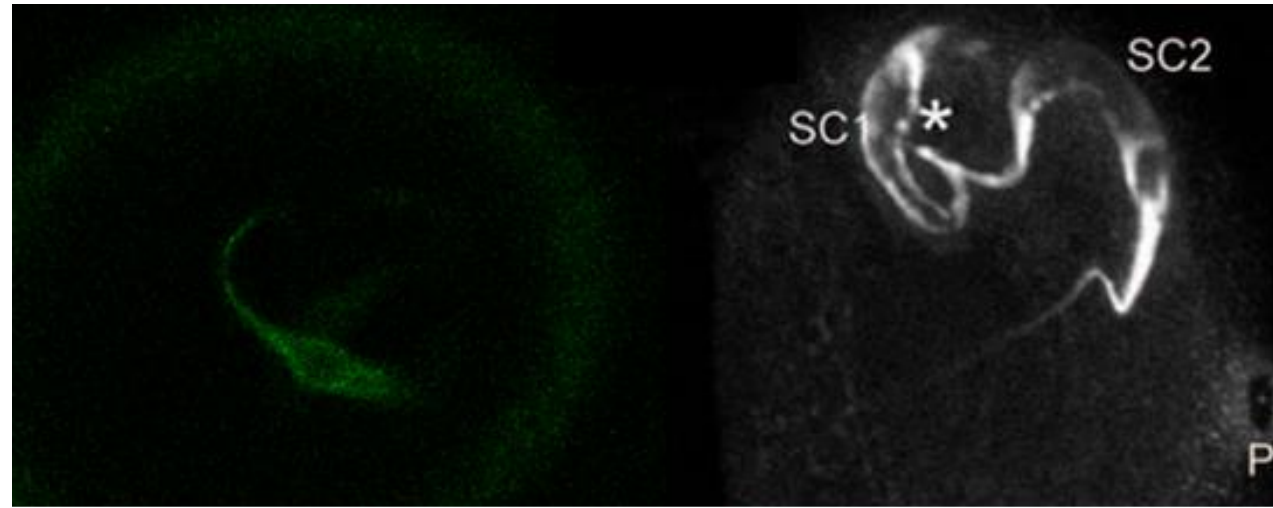
LUREs delivered by pipette



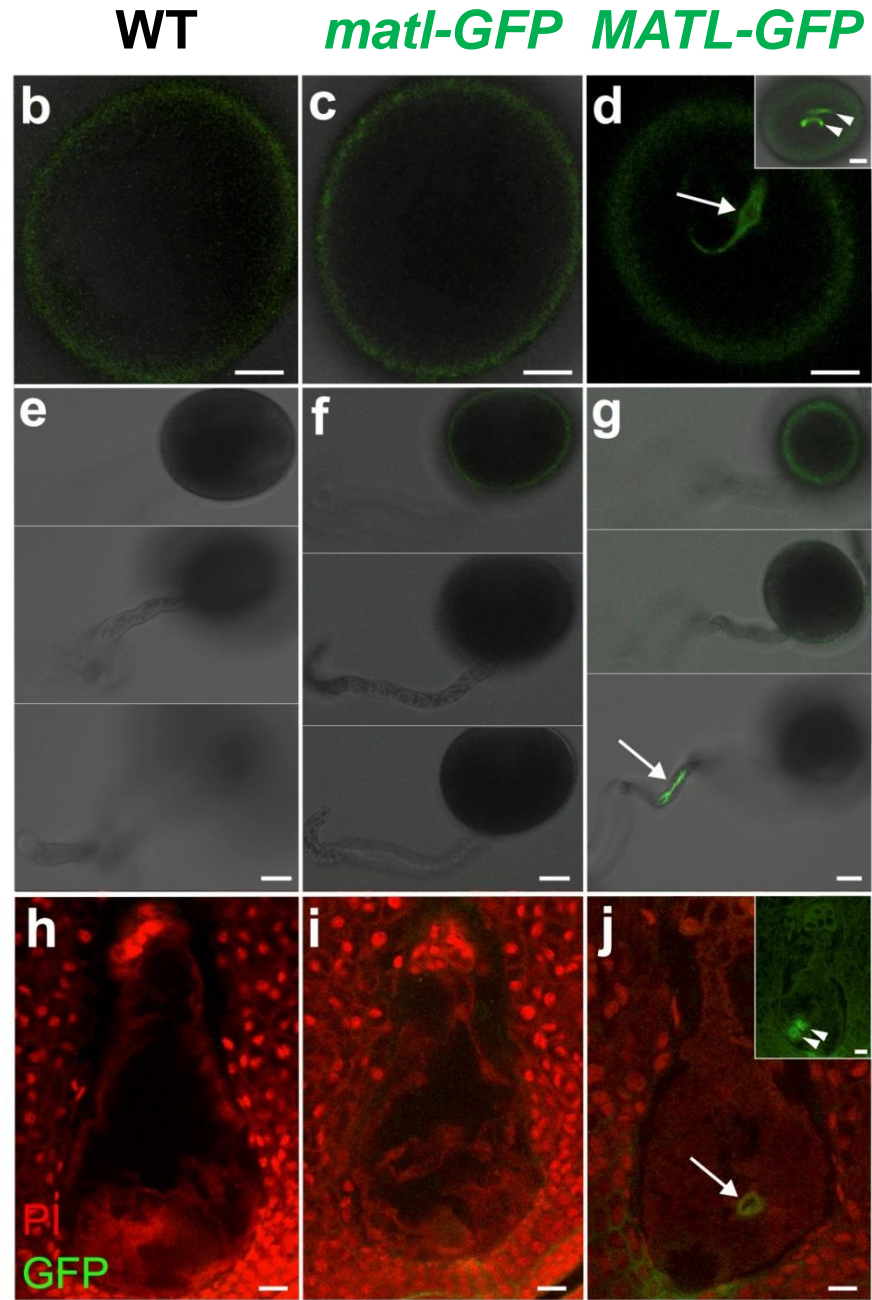
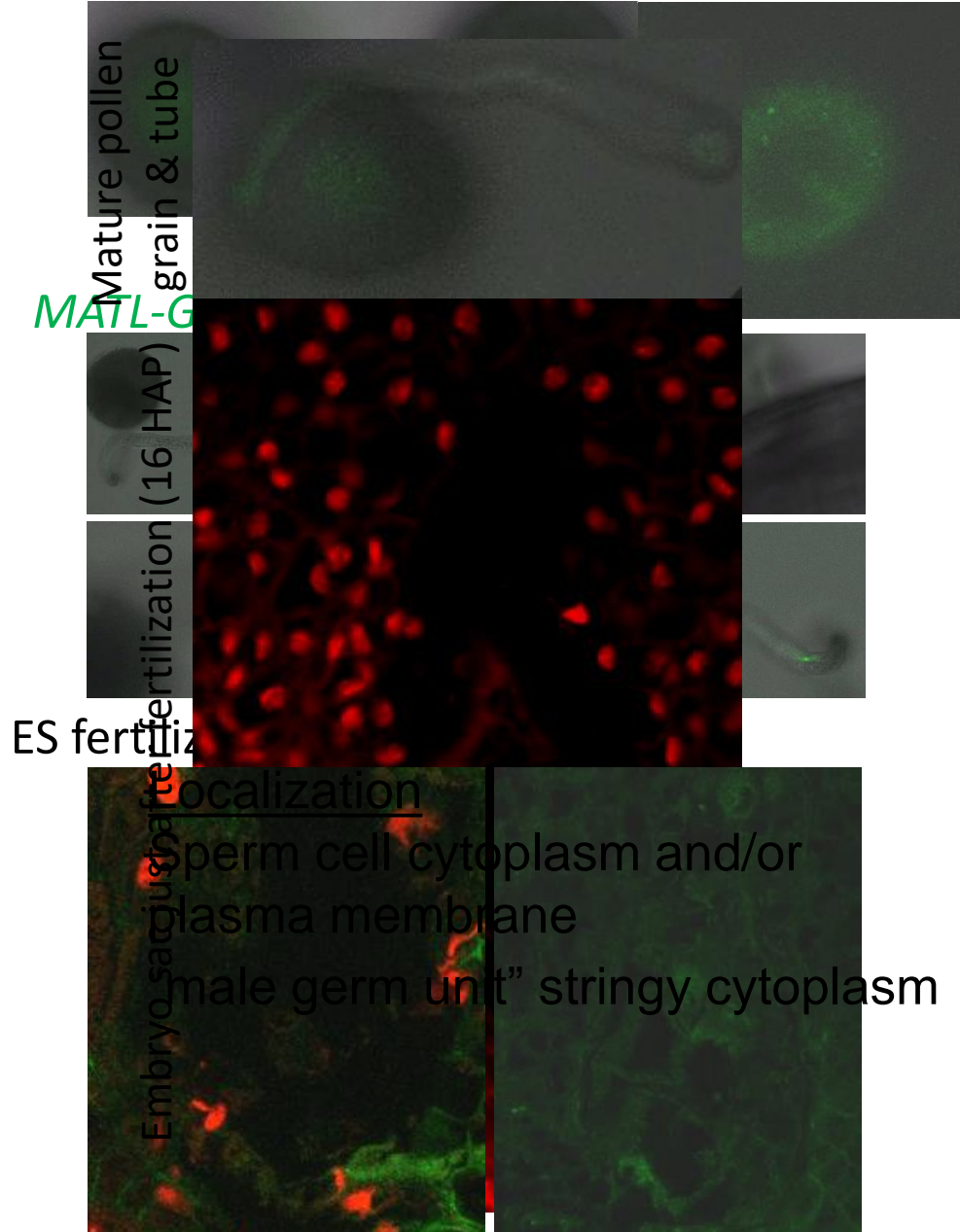
nature07882-s2 (Converted).mov

How does MATRILINEAL induce haploids?

MATL protein localizes to sperm cell membranes in pollen



matl-GFP: no signal
MATL-GFP in sperm cells (SCs)



...signal missing shortly after syngamy; not found in any maternal or zygote cell types

Mechanism --

Single nucleus sequencing reveals spermatid chromosome fragmentation as a possible cause of maize haploid induction

Xiang Li, Dexuan Meng, Shaojiang Chen, Haishan Luo, Qinghua Zhang, Weiwei Jin  & Jianbing Yan 

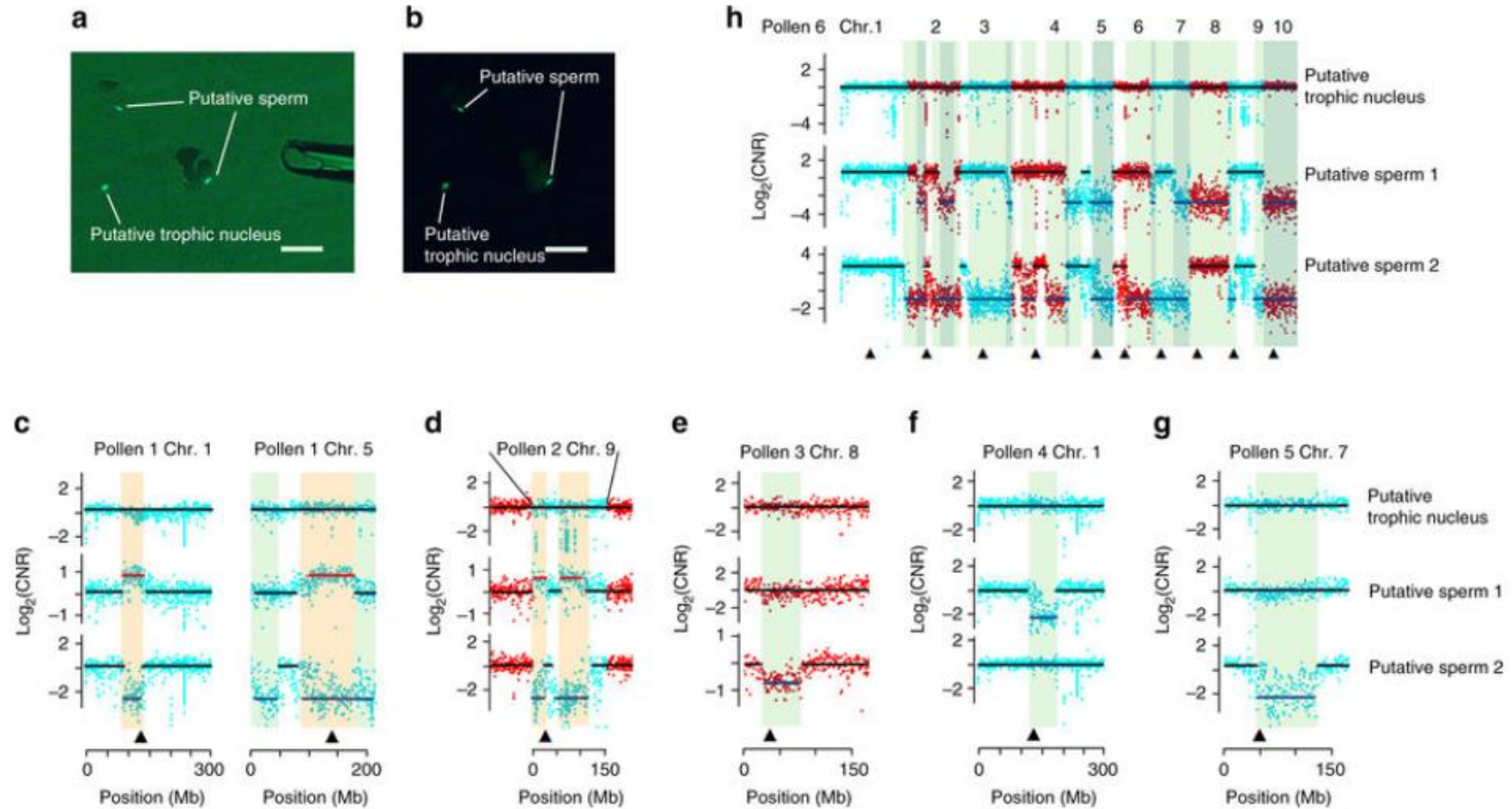
Nature Communications **8**, Article number: 991
(2017)
doi:10.1038/s41467-017-00969-8

Received: 04 January 2017

Accepted: 09 August 2017

Published online: 23 October 2017

Different sperm cells have different pieces of chromosomes missing → a sign of chromosome fragmentation



Fragmentation only in sperm cells!

Not in vegetative nucleus... suggests pollen carrying defective sperm can ride out development → fertilization may be defective or not... if not, sometimes male chromosomes will be lost in embryo afterwards

Conclusions: DNA fragmentation is progressive during late pollen development

Stage of pollen development

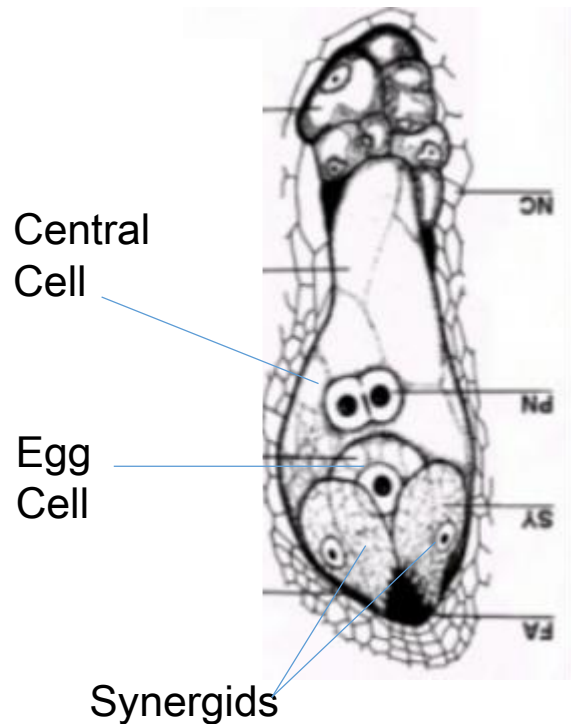
Frequency of aneuploidy

- Tetrad stage (uninucleate microspore): Few
- Mature pollen: ~15% of sperm cells

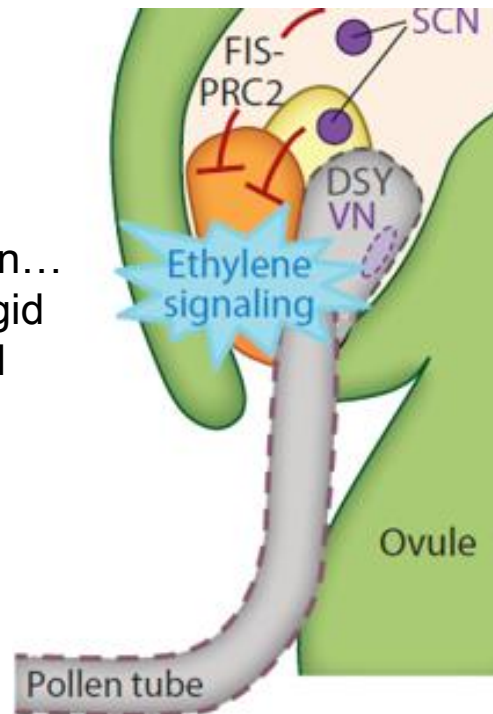
Hetero-fertilization (HF)



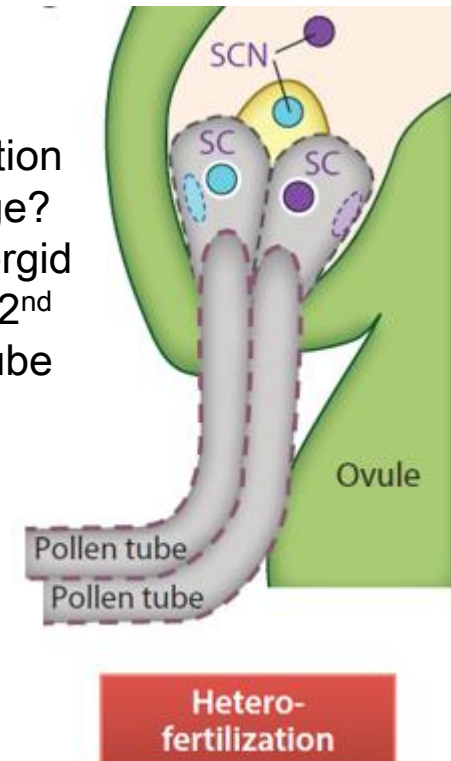
- Fertilization of egg and central cell by two different pollen grains
- Occurs very rarely in nature, but when it happens it indicates a fertilization defect



Normal fertilization...
2nd synergid is blocked



Fertilization challenge?
2nd synergid attracts 2nd pollen tube



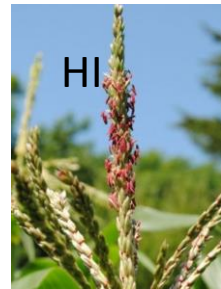
Hetero-fertilization rates are 4X higher in crosses with haploid inducer



+



- 9 & 11 am
- Both at 10
- 11 & 9 am



+



- 9 & 11 am
- Both at 10
- 11 & 9 am



+



- 9 & 11 am
- Both at 10
- 11 & 9 am

Progeny rates

~ 99 % same



~ 1 % different



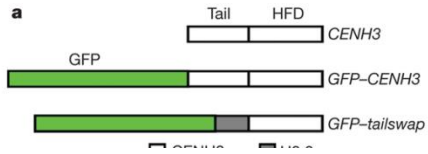
Experiments to prove post-fertilization genome elimination

- **Hyp #2: Chromosome elimination:** Male DNA is lost post-fertilization
 - Prediction: Male DNA is present in early embryo, then lost
 - Well documented in wheat x pearl millet and other wide crosses
 - Supported by evidence in literature
 - Aneuploid plants and chimeras seen after haploid induction crosses
 - Need precise tracking of male DNA markers in early embryos during HI
 - We have new data that supports this hypothesis also

Major crops
that are lacking
doubled
haploids



Arabidopsis (CENH3-tailswap) leads to haploids



- CENH3 binds centromeric DNA; helps chromosomes segregate in division
- Aberrant alleles of CENH3 produce 3-30% haploid seed (genome elimination)

Bringing Haploid Induction to new crops

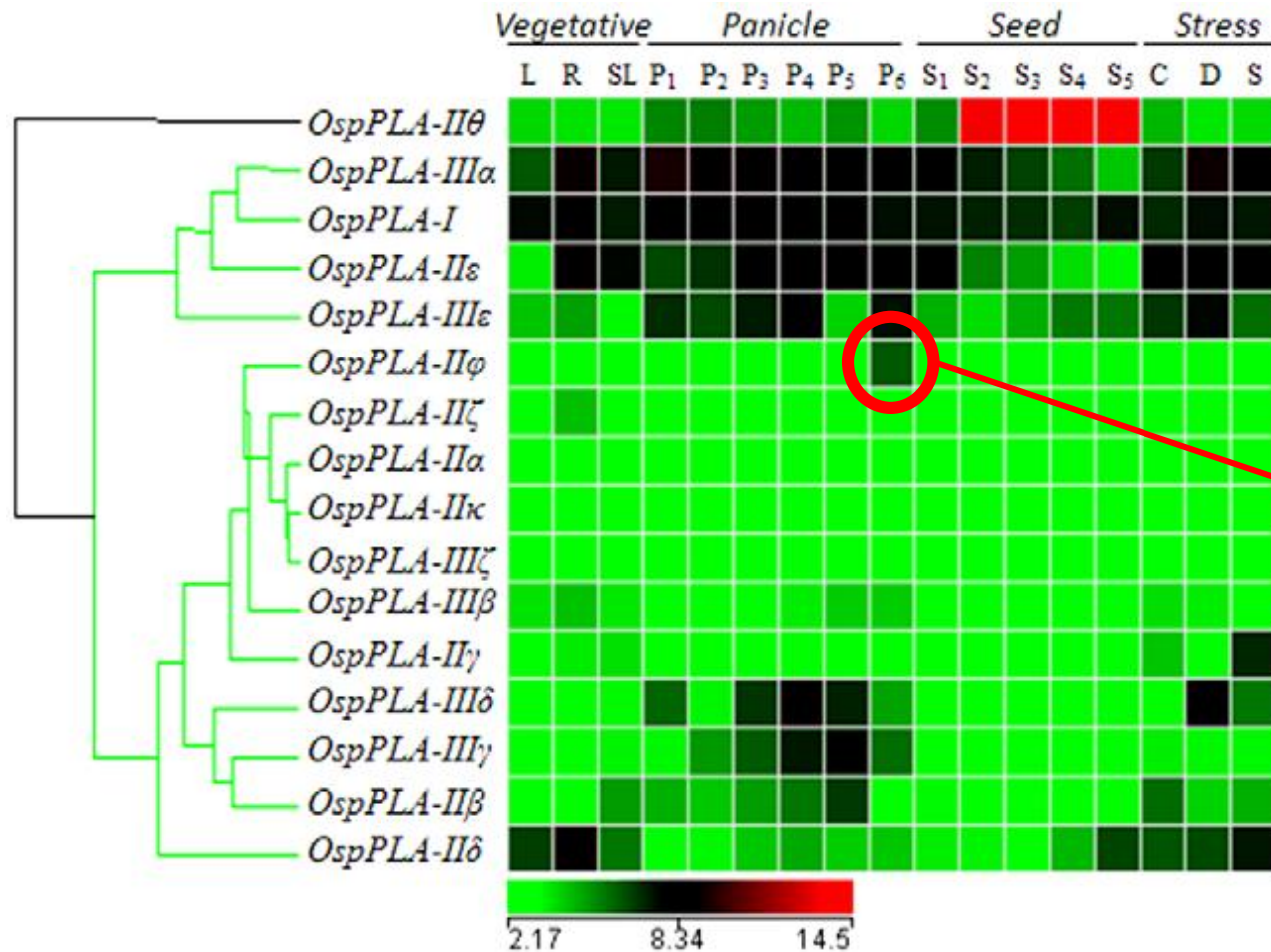
Zea mays -MAS-YSSRRPCNTCSTKAMAGSVVGEFPV-VLQQRVTVLTVDDGGVIRGLIPGTILAFLEARLQELDGFPEARLADYFDYIAGTSTGGGLITAMLTAPGKDKRPLYAAKDINHFFYMNCPRIF
 Sorghum bicolor -MATYSSRRPCNACSTKAMAGSVVGEFPV-VLQQRVTVLTVDDGGVIRGLIPGTILAFLEARLQELDGFPEARLADYFDYIAGTSTGGGLITAMLTAPGKDRRPLYAAKDINQFYMNCPRIF
 Setaria italica -MAS-YSSRRPCNACSTKAMAGSVVGEFPV-VPGQRVTVLTVDDGGVIRGLIPGTILAFLEARLQELDGFPEARLADYFDYIAGTSTGGGLITAMITTPGEDKRPPLFAARDINRFYFDNCPRIF
 Hordeum vulgare -MAS-YWCRRPCESCSTRAMAGSVVGEFPV-APGQRVTVLTVDDGGVIRGLIPGTILAFLEARLQELDGFPEARLADYFDYIAGTSTGGGLITAMLTAPGQDGRPLFAAKDVNRFFYLDNGPYIF
 Brachypodium distachyon -MAS-YACRRPCESCSTRAMAGSVVGEPT-TPGQRVTVLTVDDGGVIRGLIPGTILAFLEARLQELDGFPEARLADYFDYIAGTSTGGGLITAMITAPGEEGRPLFAAEDINRFYLDNGPQIF
 Oryza sativa v. indica -----MAGCVVGEFASAPGQRVTLTLLAIDGGGIRGLIPGTILAFLEARLQELDGFPEARLADYFDYIAGTSTGGGLITAMLAAPGDHGRPLFAASDINRFYLDNGPPIF
 Oryza sativa v. japonica MAAS-YSCRRPCEACSTRAMAGCVVGEFASAPGQRVTLTLLAIDGGGIRGLIPGTILAFLEARLQELDGFPEARLADYFDYIAGTSTGGGLITAMLAAPGDHGRPLFAASDINRFYLDNGPLIF
 Triticum aestivum -----

Zea mays PQR-SRLAAAMSALRKPKNYNGKMRSLIRSLGETRVSETLTNVIIIPAFDIRLLQPIIFSTYDAKSTPLKNALLSDVCI GTSAAPTYLPAHYFQTEDA-NGKEREYNLIDGGVAANNPTM
 Sorghum bicolor POKSSRLAAAMSALRKPKNYNGKCLRNLIIMSMLGETRVSDTLTNVVIPTFDVRLLOPIIFSTYDAKSMPLKNALLSDVCI GTSAAPTYLPAHYFQTKDAGSGKEREYNLIDGGVAANNPTM
 Setaria italica PQSRSSLAAMSALRKPKNYNGKYLRSIRSMLETRVSDALTNVVIPTFDIKLIQPIIFSTYDVKNMPLKNALLSDVCI STSAAPTYLPAHYFQIQDA-GGKREYNLIDGGVAANNPTM
 Hordeum vulgare PQRRCALAAVTASLRPRYSKYLHGKIRSMLETRVSDALTNVVIPTFDVRLLOPIIFSTYDARNMPLKNARLADICI GTSAAPTYLPAHFFHTQDD-NGKEREYNLIDGGVAANNPTM
 Brachypodium distachyon POKRSSLSVLAALTRPRYNGKFLHGKIRSMLETRVCDTLTNVVIPTFDVRLLOPIIFSTYDAKSMPLKNALLSDVCI STSAAPTYLPAHYFQTEDD-NGKEREYNLIDGGVAANNPTM
 Oryza sativa v. indica POKRCGMAAAMAALTRPRYNGKYLQKIRKMLGETRVSDTLTNVVIPTFDVRLLOPIIFSTYDAKSMPLKNALLSDICI STSAAPTYLPAHCFQTTDDATGKVFDFLIDGGVAANNPTM
 Oryza sativa v. japonica POKRCGMAAAMAALTRPRYNGKYLQKIRKMLGETRVSDTLTNVVIPTFDVRLLOPIIFSTYDAKSMPLKNALLSDICI STSAAPTYLPAHCFQTTDDATGKVFDFLIDGGVAANNPTM
 Triticum aestivum -----MLGETRLSDALTNVVIPTFDVRLLOPIIFSTYDAKSMPLKNARLADVCI GTSAAPTYLPAHFFHTHDG-NGKEREYNLIDGGVAANNPTM

Zea mays VAMTQITKQMLASKDKAEELYPVKPSNCRFLVLSIGTGSTSEQGLYTARQCSRWGI CRWLRNNGMAPIIDI FMAASSDLVDI HAAMFQSLHSDGD-YLRIQDNSLRGAAATVDAATPE
 Sorghum bicolor VAMTQITKQMLASKEKAEELYPVKPNCRKFLVLSIGTGSTSEQGLYTARQCSRWGI CRWLRNNGMAPIIDI FMAASSDLVDI HAAMFQSLHSDGD-YLRIQDNSLRGAAATVDAATPE
 Setaria italica VAMTQITKQMLAKDK--EELYPVKPEDCRKFLVLSIGTGSTSEGLFTARQCSRWGVVWRWLRNNGMAPIIDI FMAASSDLVDI HAAMFQSLHSDG-----HSLRGAAATVDAATPE
 Hordeum vulgare VMTIQITKQMVKDR--EELYPVKPSDCGKFLVLSIGTGSTSDQGLYTAKQCSQWGI IRWLRNNGMAPIIDI FMAASSDLVDI HAAMFQSLHSDGN-YLRIQDNSLRGPAATVDAATPE
 Brachypodium distachyon VAMTQITKQIMAKDK--EELYPVKPSDCGKFLVLSIGTGSTSDQGLYTAKQCSRWGI IRWLRNNGMAPIIDI FMAASSDLVDI HAAMFQSLHSDGDCYLRIQDNSLRGAAATVDTATPD
 Oryza sativa v. indica VAMTQITKQIMVSKDK--EELYPVKPSDCGKFLVLSIGTGSTSDQGLYTARQCSRWGI VWRWLRNNGMAPIIDI FMAASSDLVDI HAAMFQSLHSDGD-YLRIQDNTLHGDAATVDAATRD
 Oryza sativa v. japonica VAMTQITKQIMVSKDK--EELYPVKPSDCGKFLVLSIGTGSTSDQGLYTARQCSRWGI VWRWLRNNGMAPIIDI FMAASSDLVDI HAAMFQSLHSDGD-YLRIQDNTLHGDAATVDAATRD
 Triticum aestivum VAMTQITKQMGKDR--EELYPVPEPSDCGKFLVLSIGTGSTSDQGLYTAKQCSQWGI ISWLRNNGMAPIIDI FMAASSDLVDI HAAMFQSLHSDAN-YLRIQDNSLRGPAATVDAATPE

Zea mays NMRITLVGIGERMLAQVSRVNVETGRYEPVTEGGSNADALGGLARQLSEERRRLARRVSAINFRGS----RCAS--YDI
 Sorghum bicolor NMRITLVGIGERMLAQVSRVNVETGRYEPVTEGGSNADALAGIARQLSEERRRLARRVSAIVSSGGASRRTCASKVSNV
 Setaria italica NMRITLVGIGERMLAQVSRVNVETGRYEPVTEGGSNADALVALARQLSDERRRARRARRAAACAGGS----RCCSP-VKT
 Hordeum vulgare NMARELLRIGERMLAQVSRVNVETGRYEEIRGAGSNADALAGFAQLSDERRRRLGRRRVGAGRLKS----RR-----
 Brachypodium distachyon NMRALVIGERMLAQVSRVNVETGRYEEVPGAGSNADALAGFAQLSDERRRRLGRRRVGAGRLKS----RC-----
 Oryza sativa v. indica NMRALVIGERMLAQVSRVNVETGRYEEVPGAGSNADALRGFARQLSEERRRRLGRRRNA CGGGEG----EPGCVACKR
 Oryza sativa v. japonica NMRALVIGERMLAQVSRVNVETGRYEEVPGAGSNADALRGFARQLSEERRRRLGRRRNA CGGGEG----EPGCVACKR
 Triticum aestivum NMARELLRIGERMLAQVSRVNVETGRYEEVKGAGSNADALAGFAQLSDERRRRLGRRRVGAGRLKS--SR-----

OsPLA ψ is the closest homolog in rice

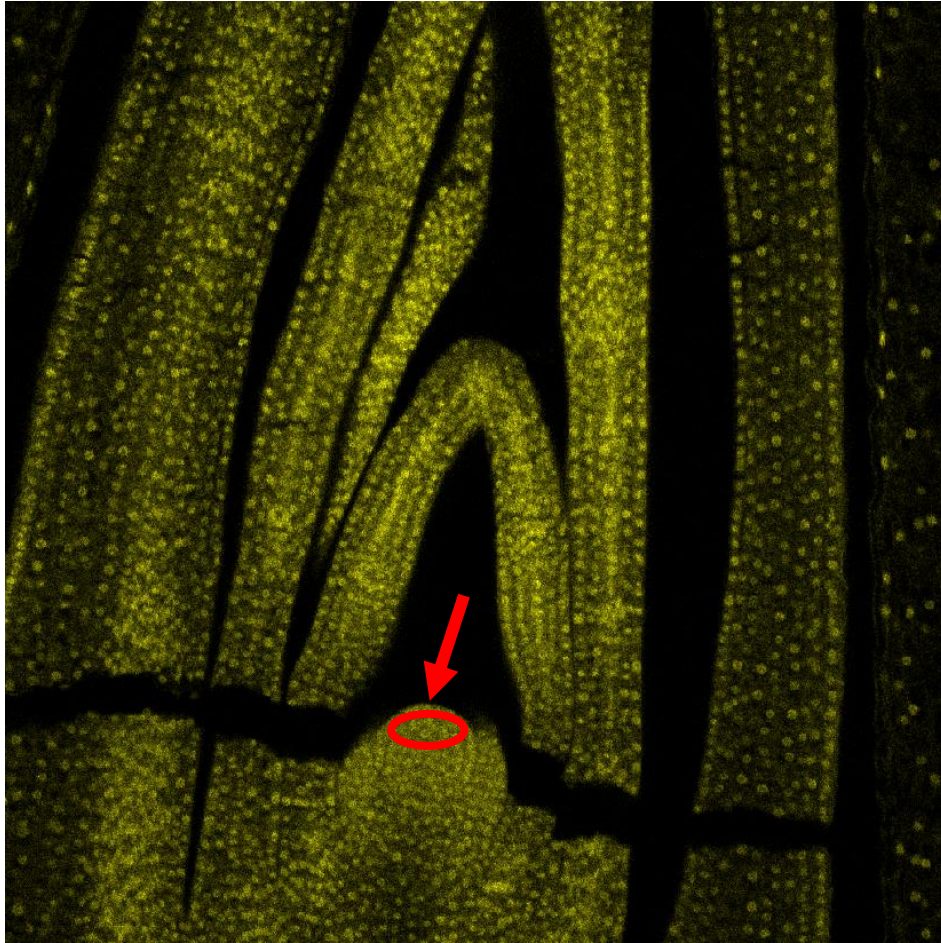
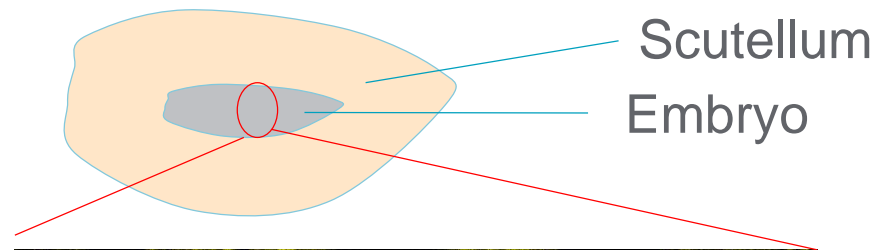


Os03g27610

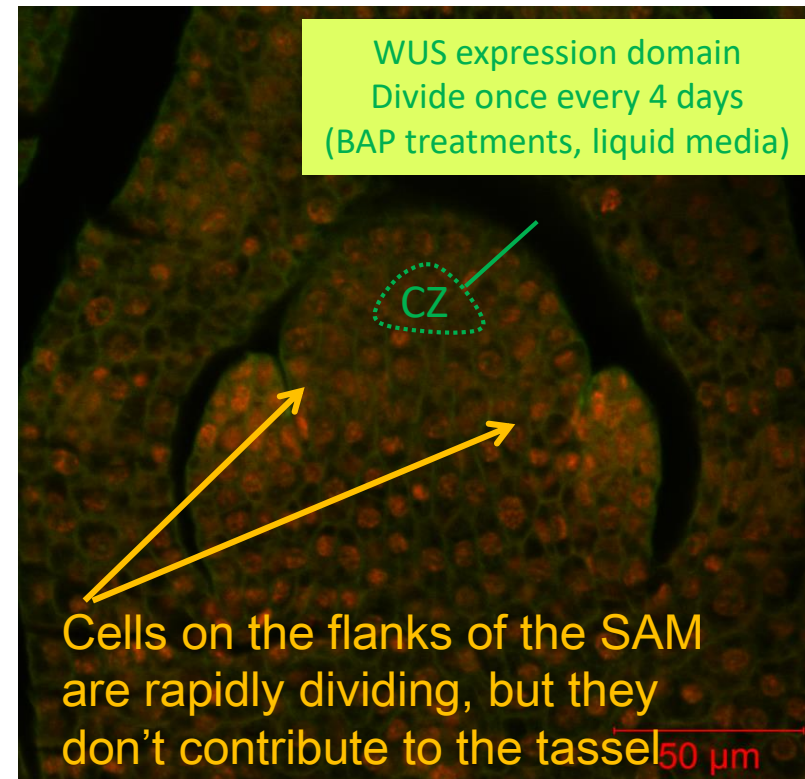
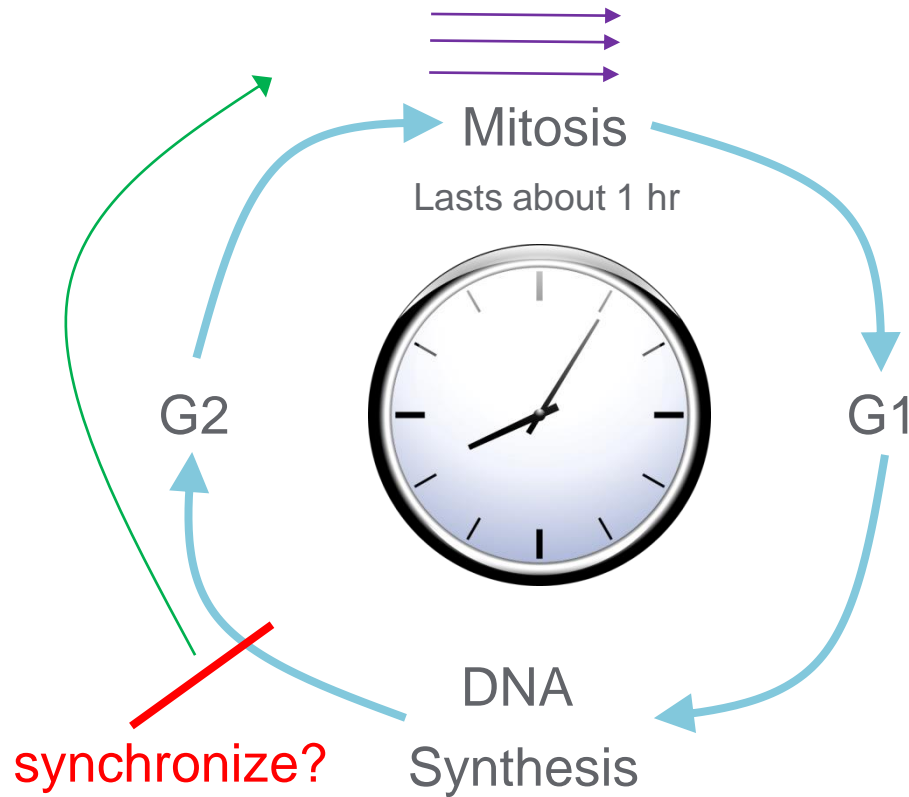
Closest homolog
to *MATL*

Also pollen-specific

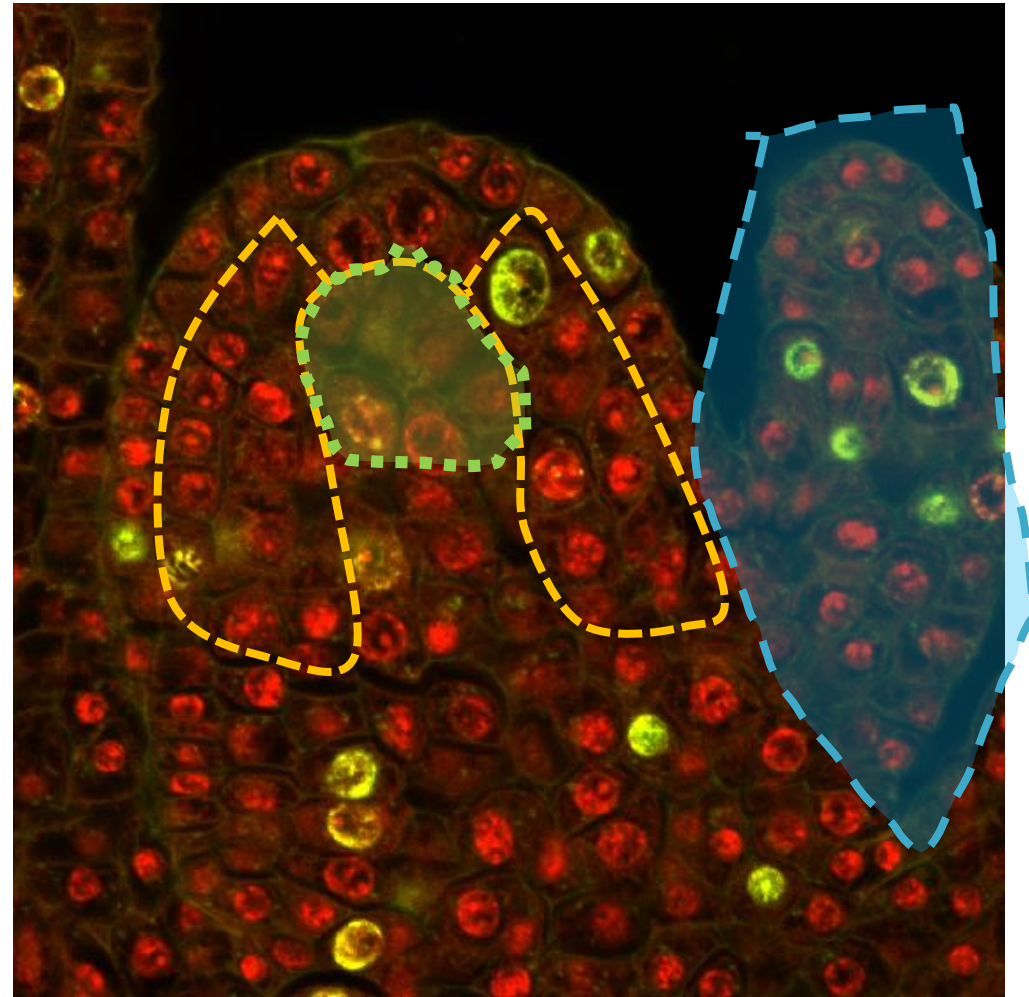
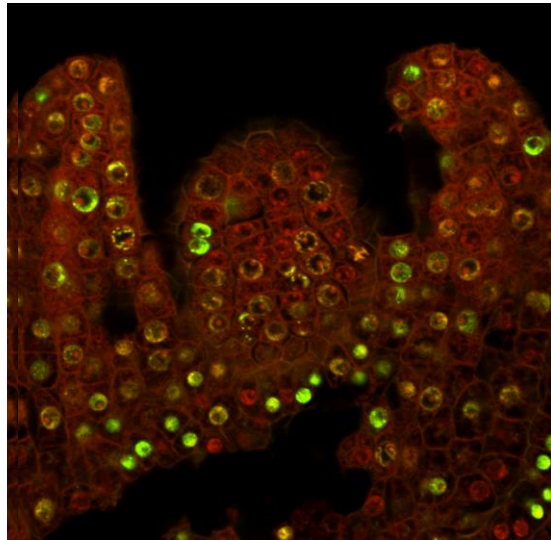
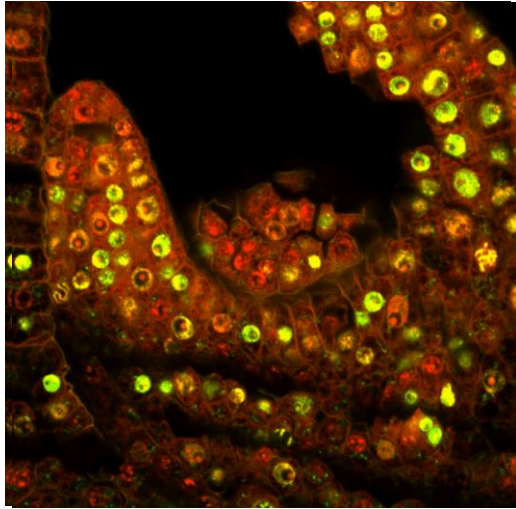
**Synchronize the 4-8 meristem cells
that → give rise to pollen**



Find treatments to boost / synchronize cell division rate of SAM CZ cells



EdU: Incorporated into dividing cells during DNA synthesis



Acknowledgements

- Syngenta Seeds Research, Reproductive Biology Team
 - **Dakota Starr, Selena Tran, Laura Hamon, Mason McNair, Erin Burch, Guozhu Tang, Jacobo Rozo Posso**
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- Maize transformation: **Lee Richbourg, Zhongying Chen, Shujie Dong, Siva Elumalai, David Bradley, Anna Prairie, Heng Zhong, Sam Nalapalli**
- Performance Traits: **Ian Jepson, Dave Skibbe, Erik Dunder, Paul Bullock**
- **Chris Leming, Stacy Miles**

Research Triangle Park site founded in 1984 by Dr. Mary-Dell Chilton – Queen of Agrobacterium



* Clive James, 2012

- Led research study in '70s and '80s that resulted in the first biotech plants
- With Dr. Chilton's technical leadership, Syngenta was the first company to commercialize a biotech trait in corn
- Dr. Chilton named 2013 World Food Prize laureate. In 2015, inducted to the National Inventors Hall of Fame, USDA Hall of Heroes and National Academy of Inventors

Key R&D centers across the world

Unrivalled global breadth



The Syngenta RTP Innovation Center

- Phase I: Construction of the Advanced Crop Lab started Q4/2011
 - Construction completed Q1/2013
 - Investment of \$72 million
- Phase II: Construction of the RTP Innovation Center started Q4/2013
 - Construction completed Q2/2016
 - Investment of \$94 million



The Syngenta RTP Innovation Center: lab and office wings

- Lab space (136,000 sq. ft. total)
 - 100,000 sq. ft. of labs
 - 36,000 sq. ft. of offices and collaboration spaces
 - Flexible, fit-for-purpose, open laboratory environments
 - State-of-the-art fermentation suite



- Office space
 - 64,000 sq. ft.
 - Multi-workplace office environments to enable interaction between disciplines
- Total Innovation Center
 - 200,000 sq. ft.
- Houses approximately 475 employees.
 - 375 Syngenta employees
 - 100 resident contractors.

The Syngenta RTP Innovation Center: Advanced Crop Lab

- 340,000 sq. ft. of State-of-the-art facilities

The Advanced Crop Lab (140,000 sq. ft.)

Under Glass: 40,000 sq. ft.

Headhouse + Chambers: 35,000 sq. ft.

Support Areas: 65,000 sq. ft.



- **Greenhouses**

- Positioned to receive optimal sunlight
- Made of low carbon glass with antireflective coating and prismatic haze patterns.
- Independently controlled room temperature and humidity
- Control of day light exposure and gases such as carbon dioxide

- **Growth Chambers**

- Designed specifically for Syngenta using technology developed by the International Space Program to study the impact of different atmospheres on crop performance.
- Ability to precisely control all environmental variables (light, temperature, moisture, etc.)
- Ability to precisely measure plant responses